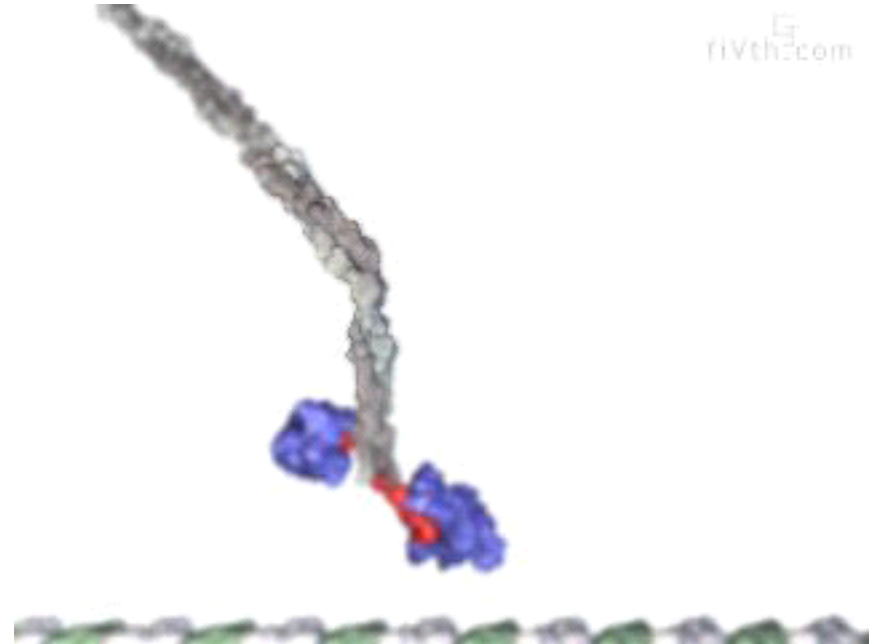
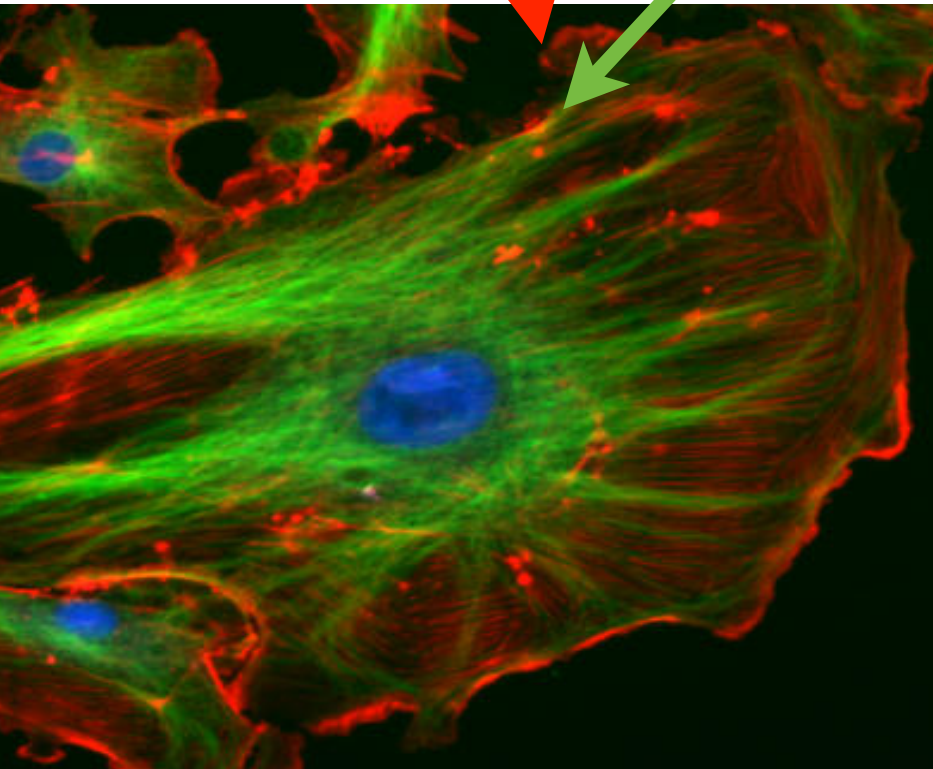
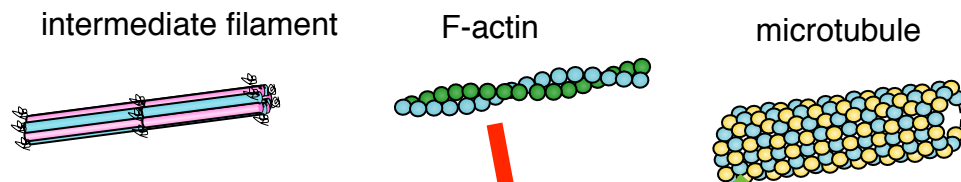


The cytoskeleton, an active *solid*

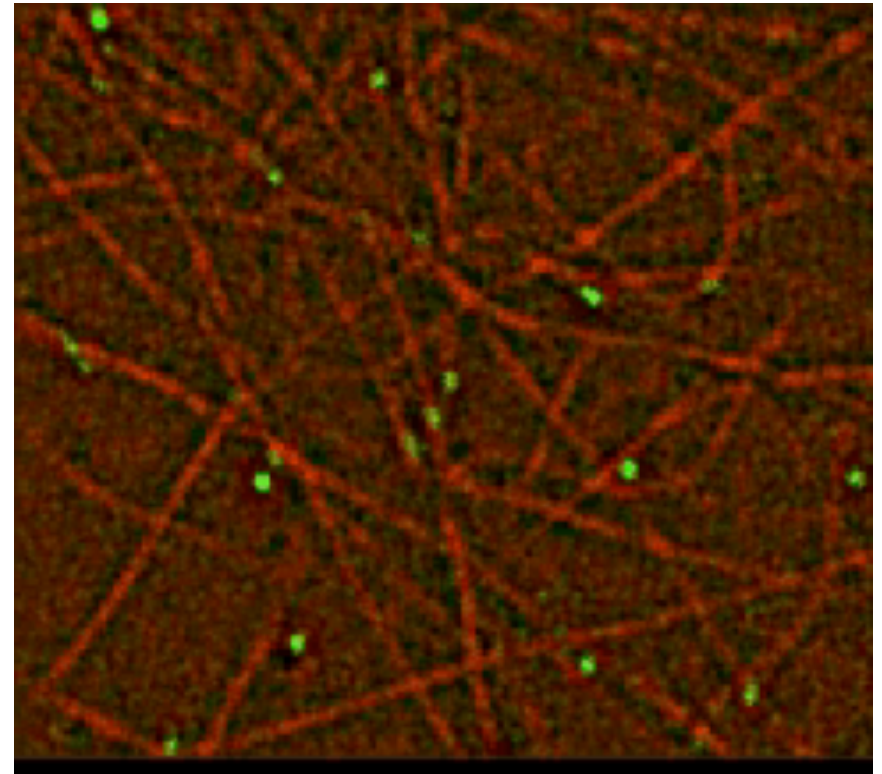
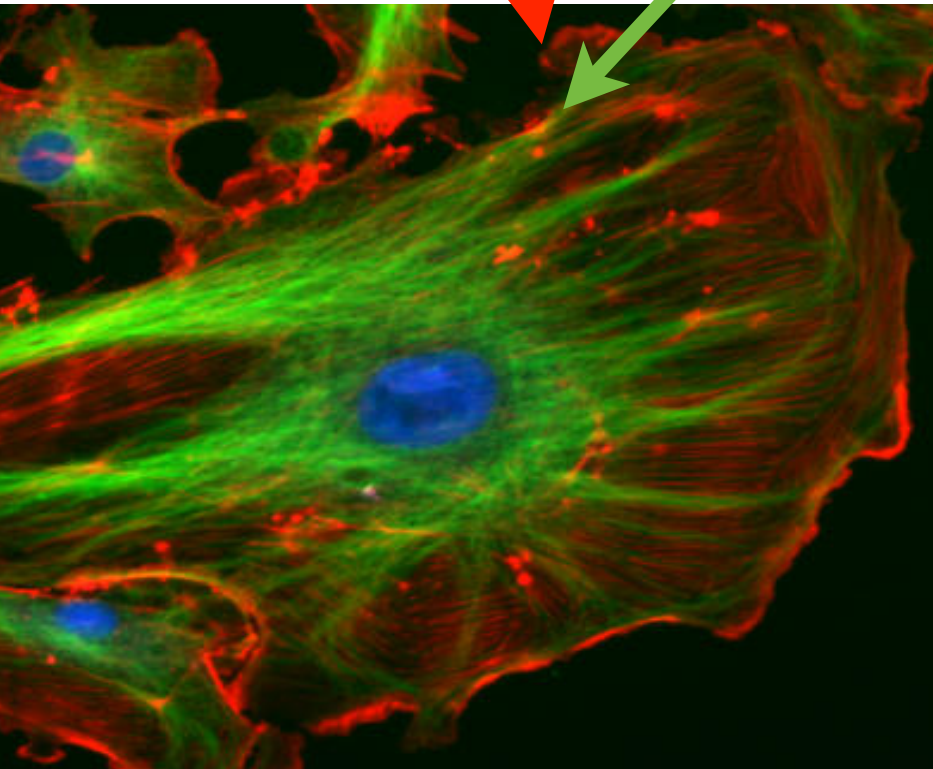
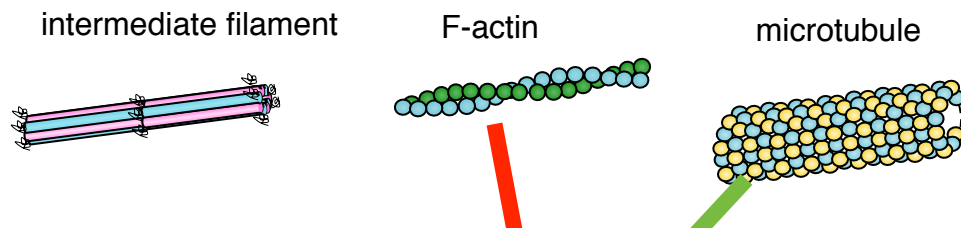
The Cell



Vale lab, *UCSF*

The cytoskeleton, an active *solid*

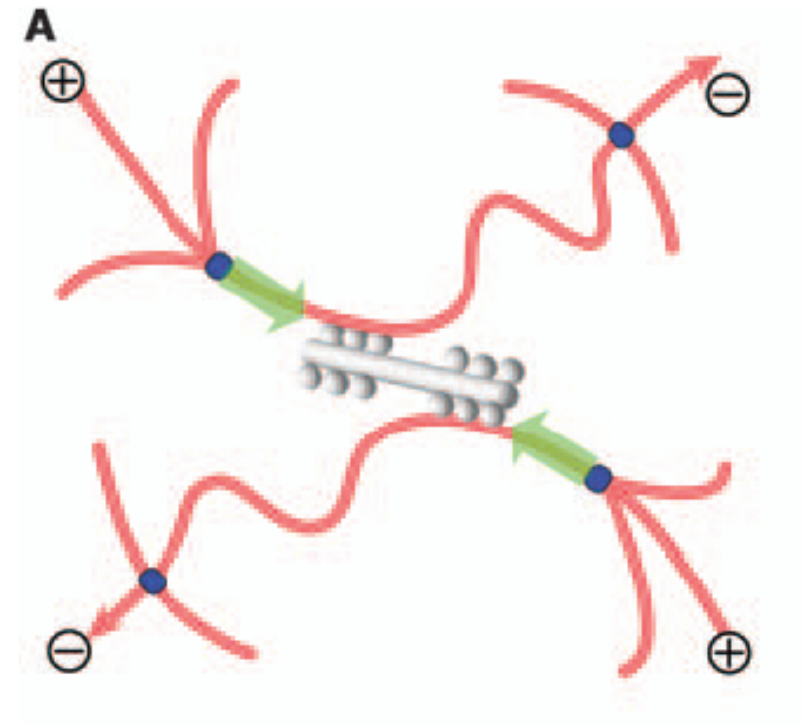
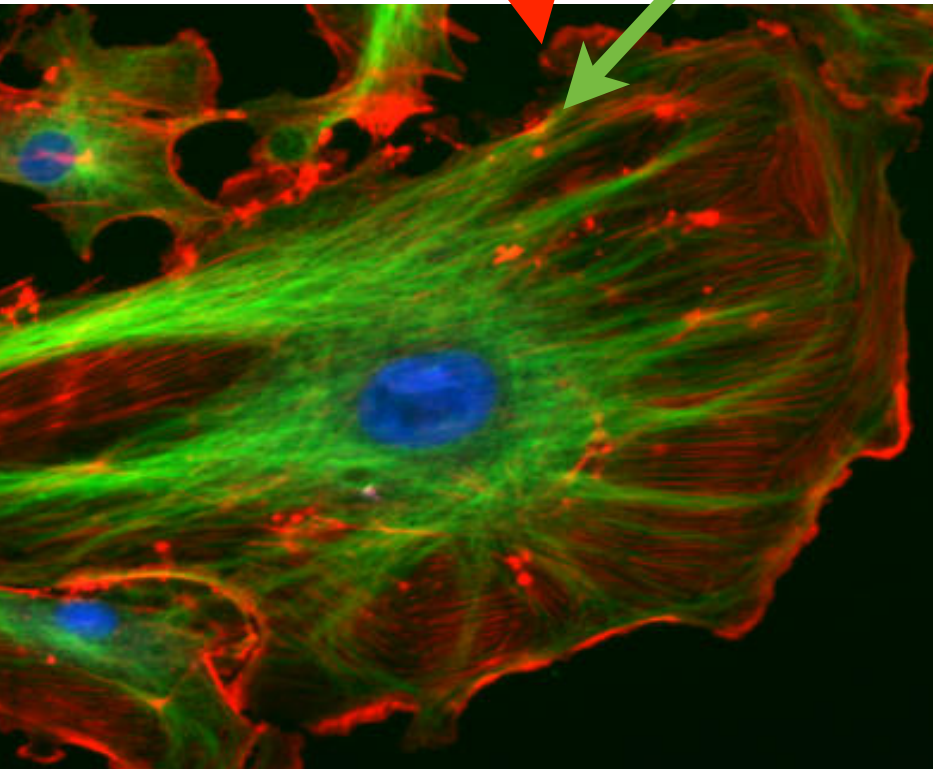
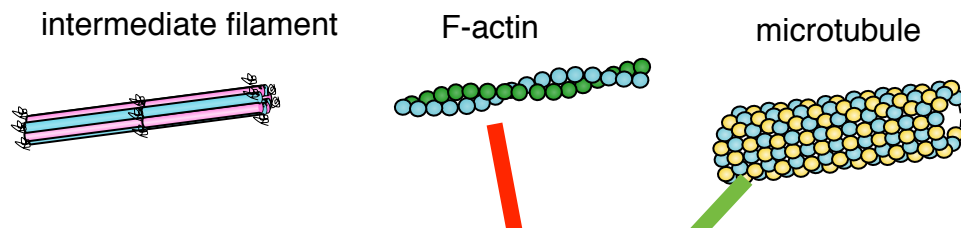
The Cell



with Kapitein et al.,
Current Biol., (2010).

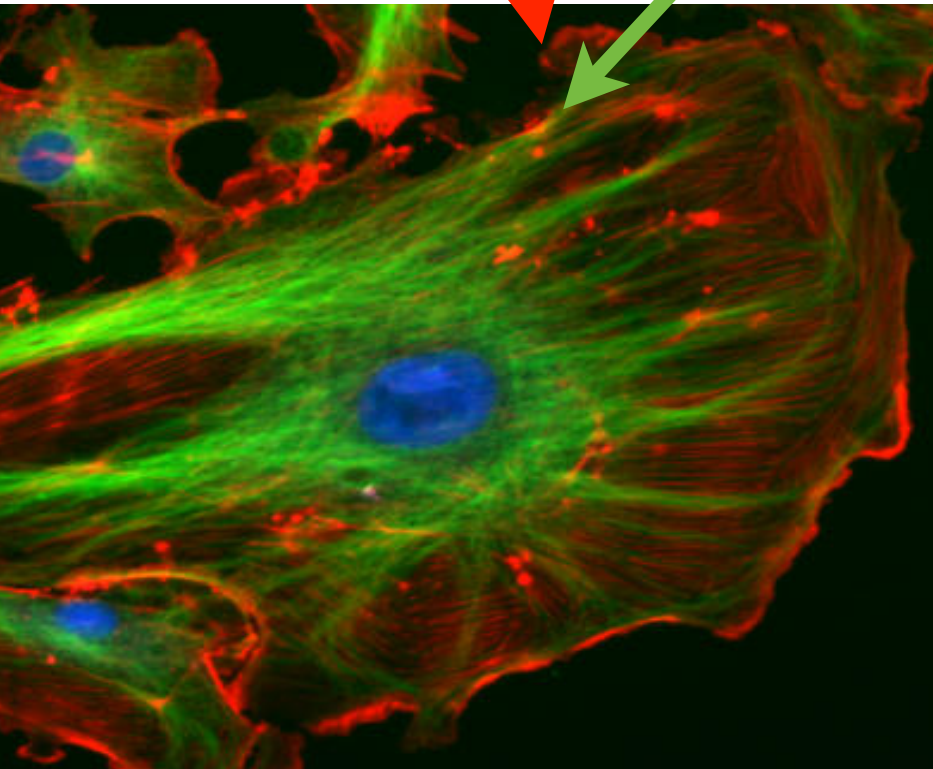
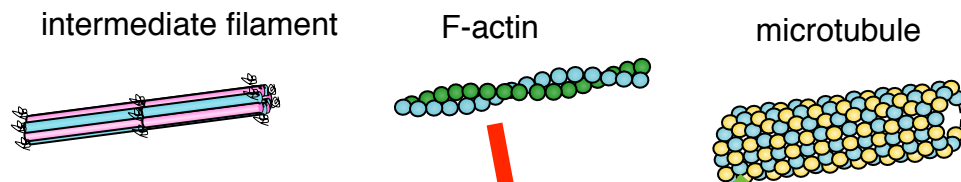
The cytoskeleton, an active *solid*

The Cell

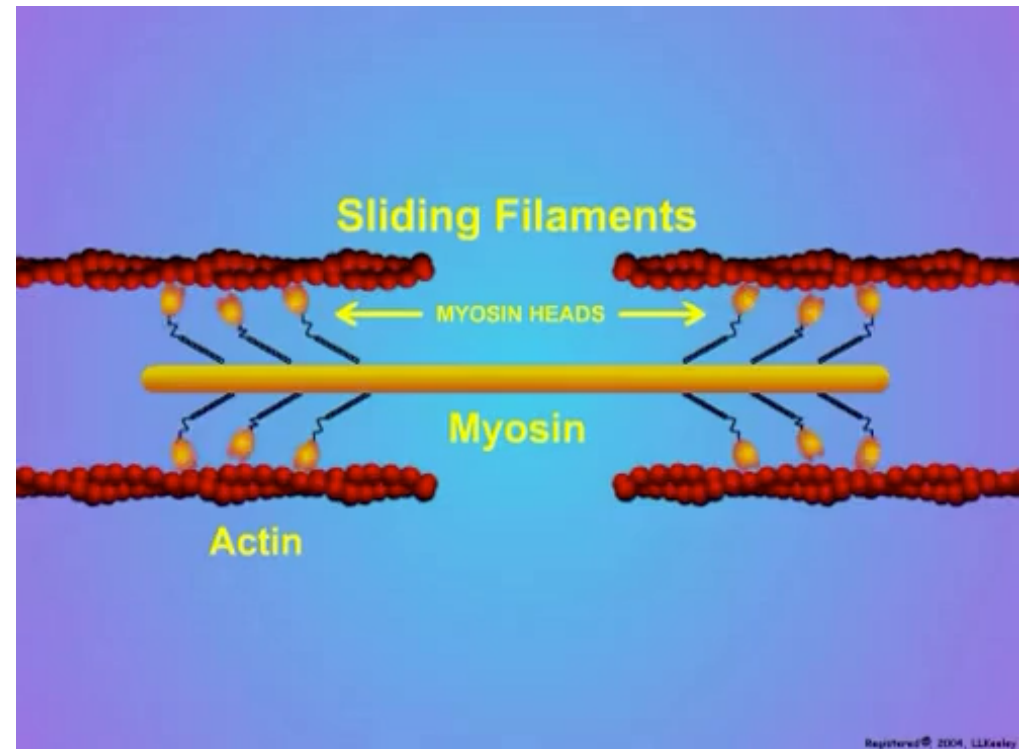


The cytoskeleton, an active *solid*

The Cell



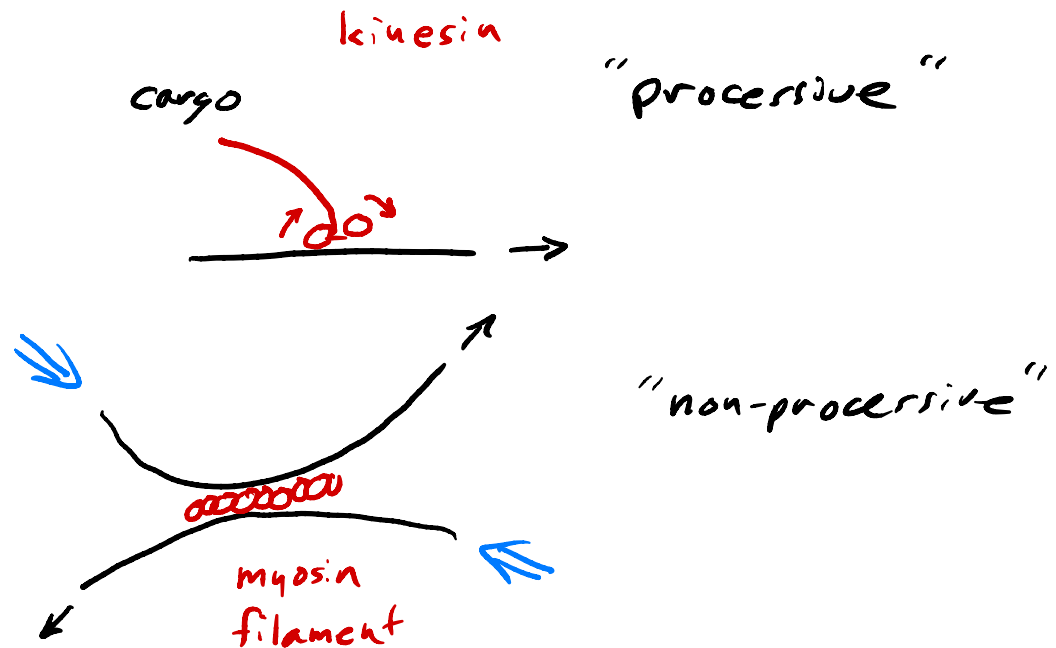
Myosin in muscle



https://www.youtube.com/watch?v=zQocsLRm7_A&feature=youtu.be

Molecular motors

- Linear motors



- Rotary Motor



Biopolymers & their networks in cells

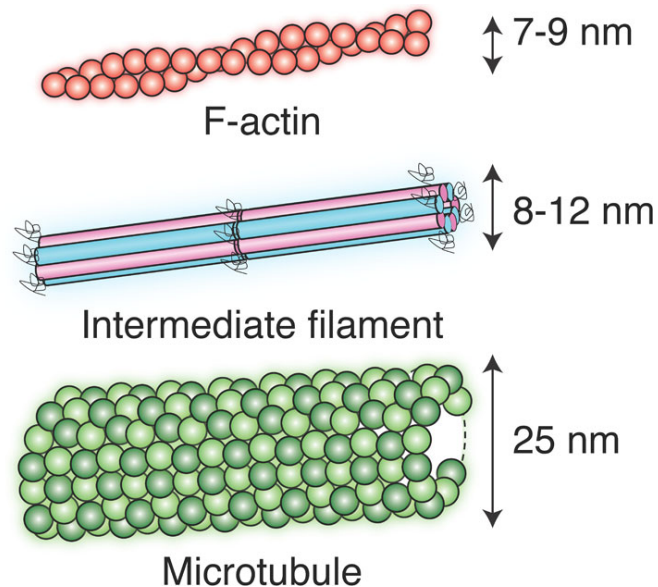


FIG. 4 (color online). The three families of cytoskeletal filaments, including F-actin, intermediate filaments, and microtubules.

**see, e.g., Broedersz and FCM,
Rev Mod Phys, 86: 995 (2014).**

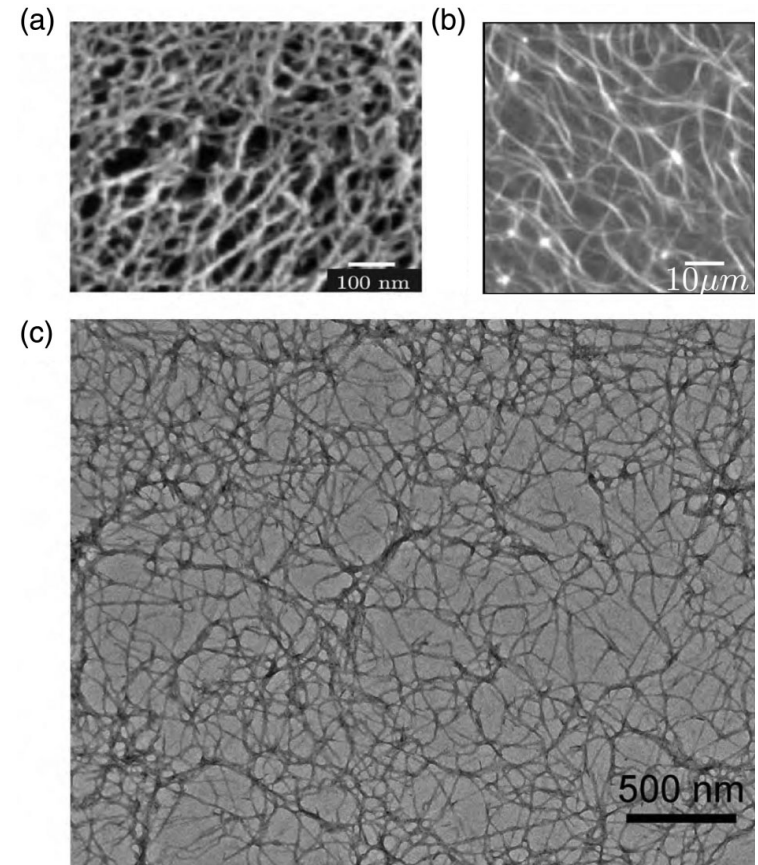
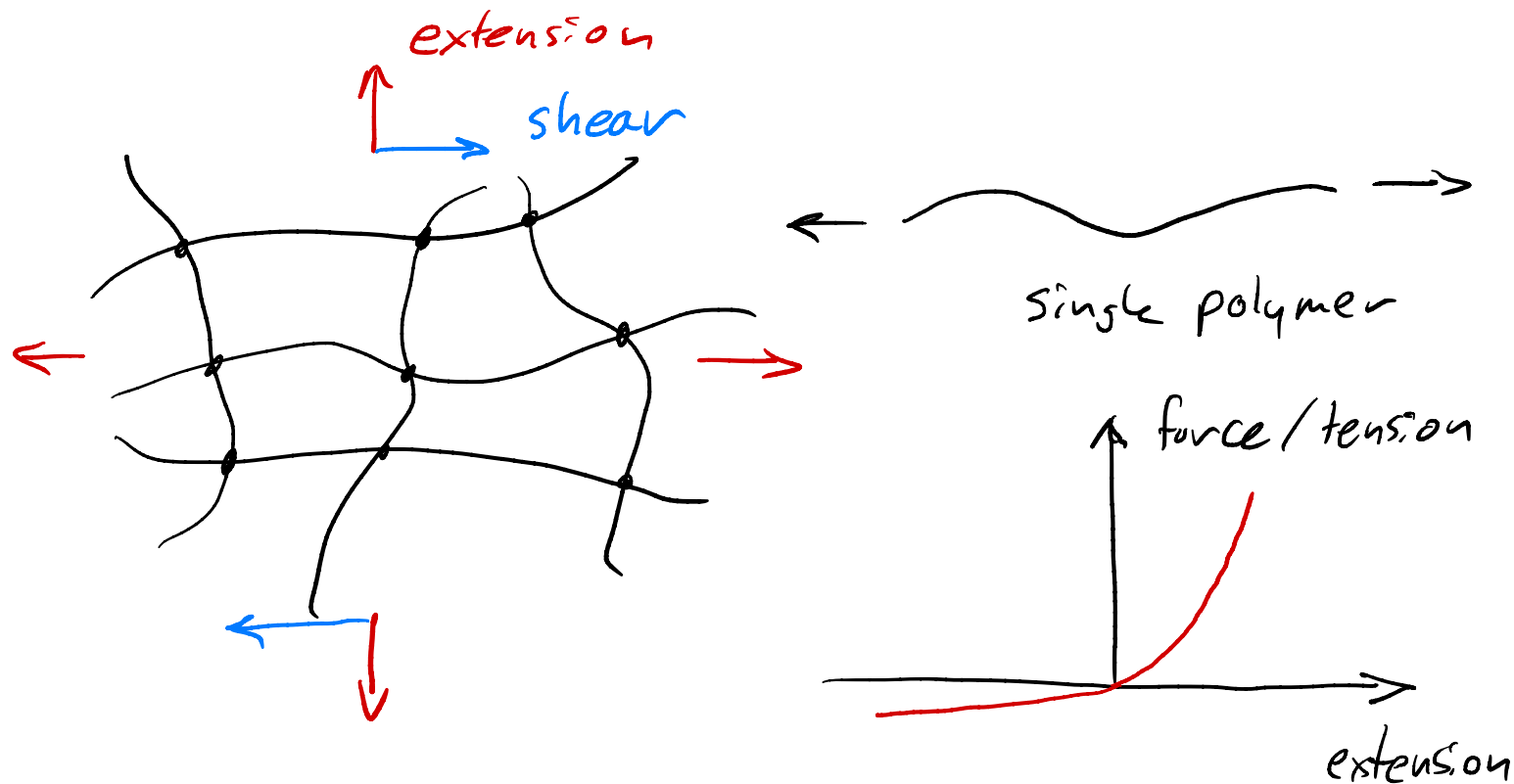


FIG. 3. (a) Electron micrograph of a fixed and rotary-shadowed filamin-F-actin network at an actin concentration 1 mg/ml, average filament length 15 μm , and a filamin:actin molar ratio of 0.005:1. From [Kasza *et al.*, 2009](#). (b) Confocal microscopy image of a fluorescently labeled bundled filamin-F-actin network at high filamin concentrations. From [Kasza *et al.*, 2010](#). (c) Electron micrograph of a fixed and rotary-shadowed Vimentin network. Courtesy of Y-C. Lin and D. Weitz (Harvard).

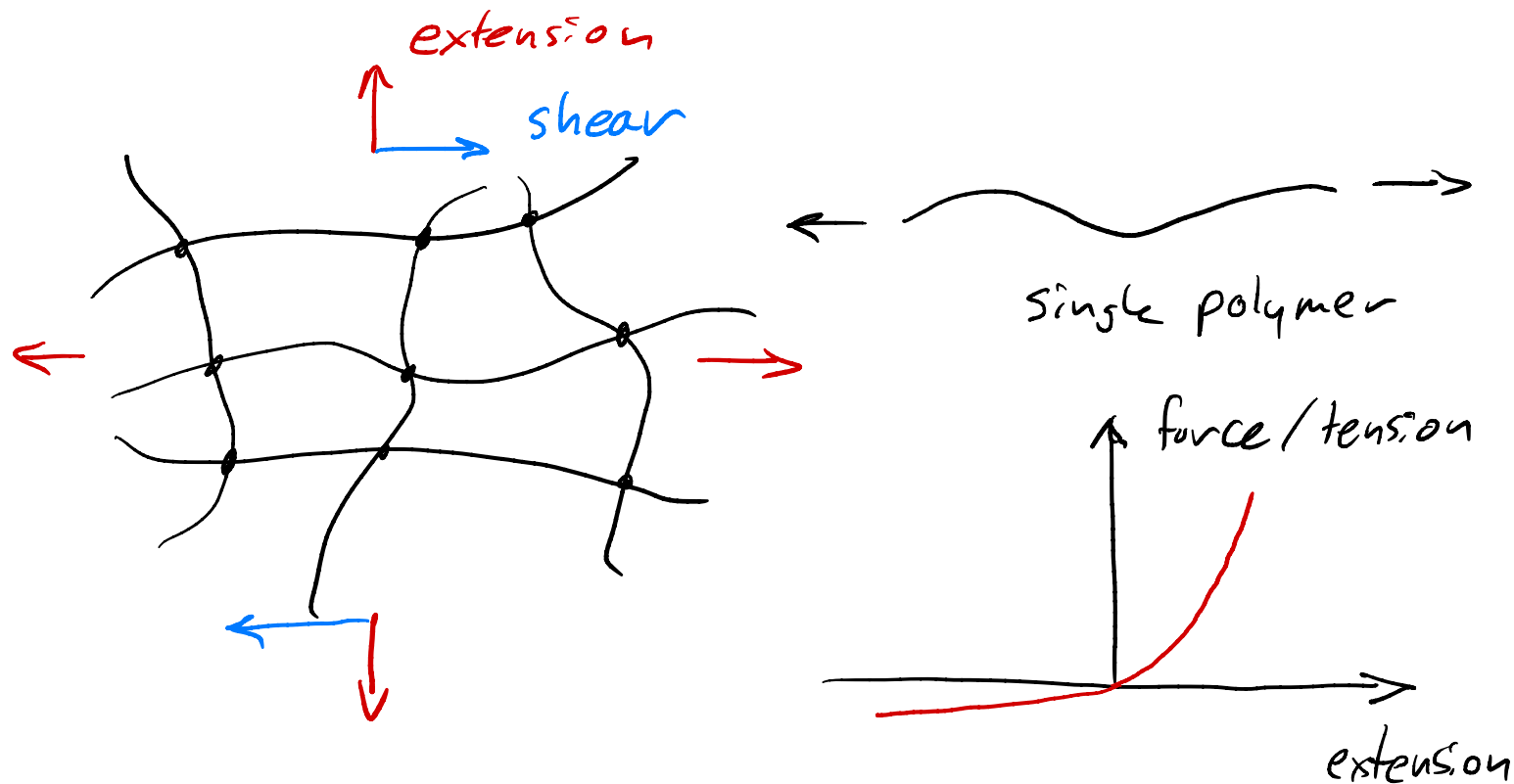
Networks stiffen under extension/deformation

Networks of biopolymers



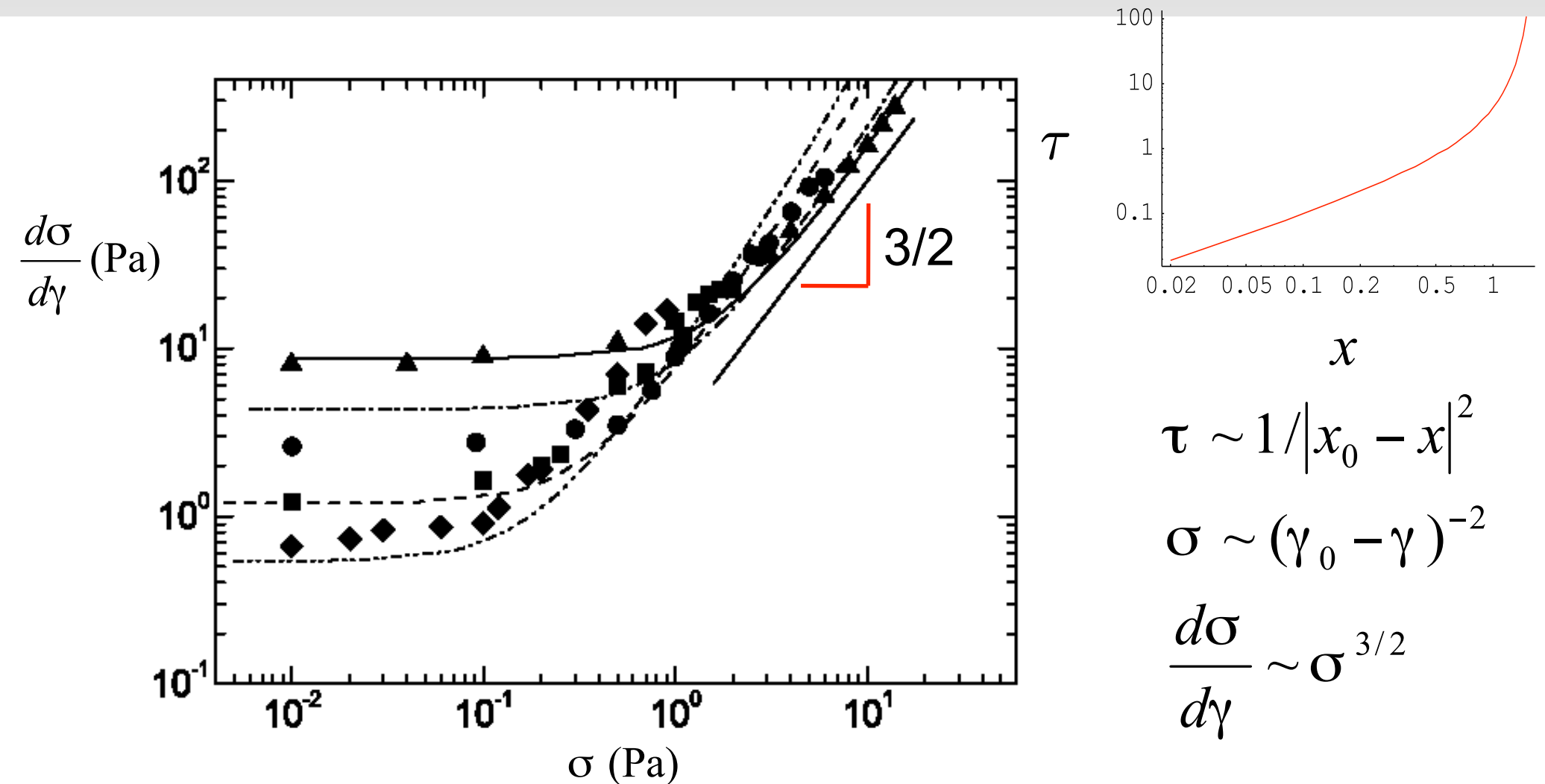
Networks stiffen under extension/deformation

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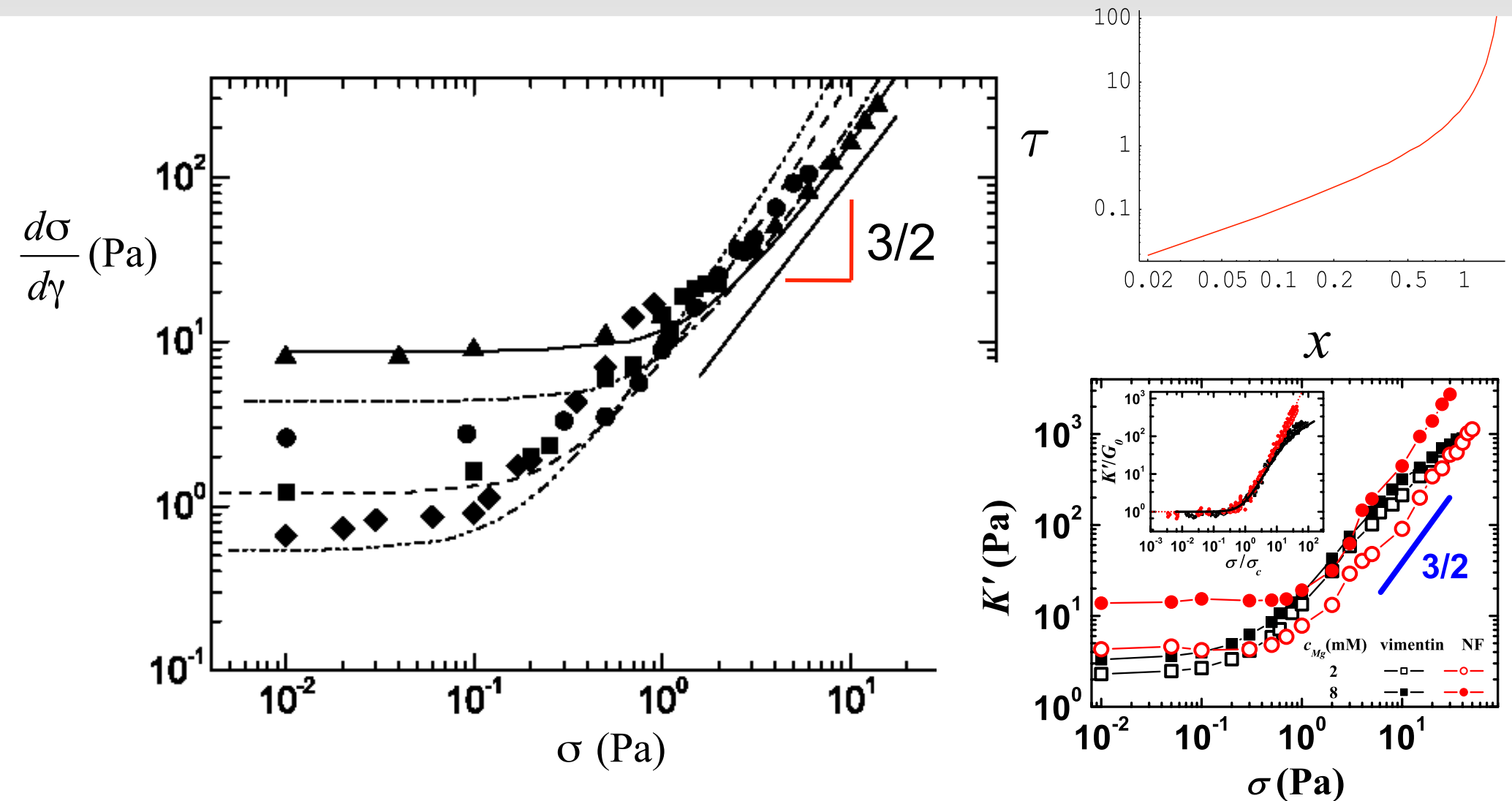
Scaling of Stress Stiffening



Quantitative *actin Network* elasticity from *single filament* stat. mech.

Gardel, et al., *Science* (2004) & *PRL* (2004); Lin, Yao, et al. *PRL* & *Biophys J* 2010.

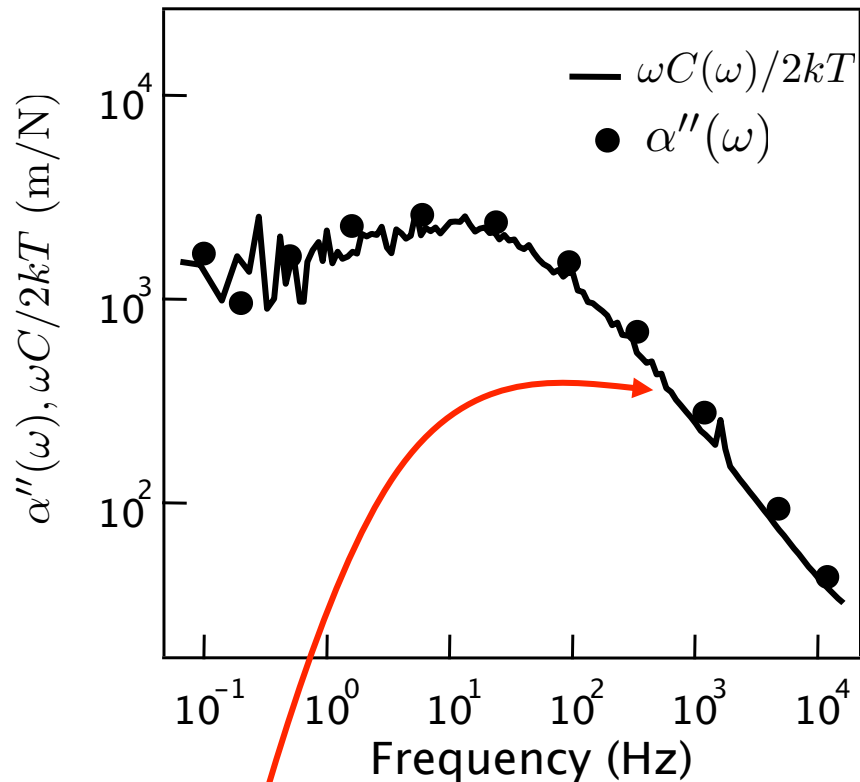
Scaling of Stress Stiffening



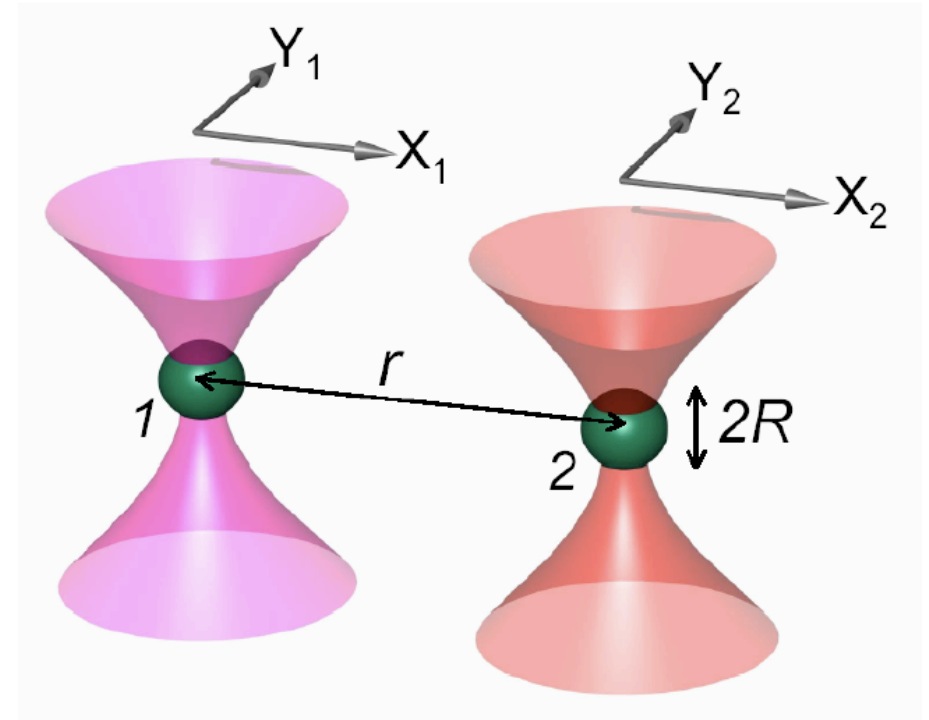
Quantitative *actin Network* elasticity from *single filament* stat. mech.

Gardel, et al., Science (2004) & PRL (2004); Lin, Yao, et al. PRL & Biophys J 2010.

Microrheology and thermal motion

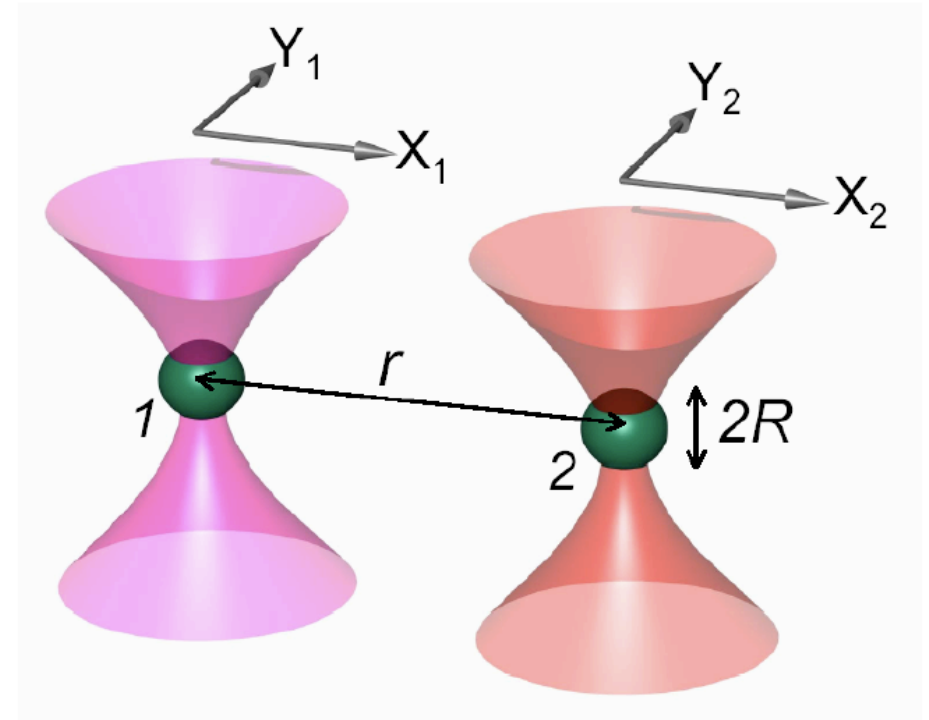
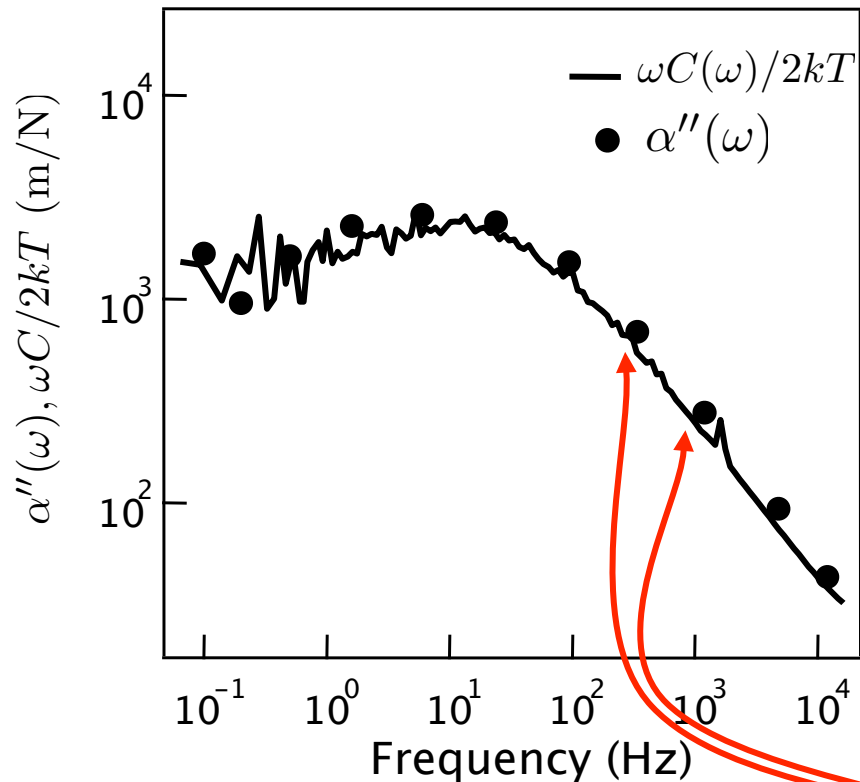


$$C(\omega) = \int \langle x(t)x(0) \rangle e^{i\omega t} dt = \frac{2kT}{\omega} \alpha''(\omega)$$



Simultaneous measurement of **passive** fluctuations and active response to calib. forces (Mizuno, et al. 2007)

Microrheology and thermal motion



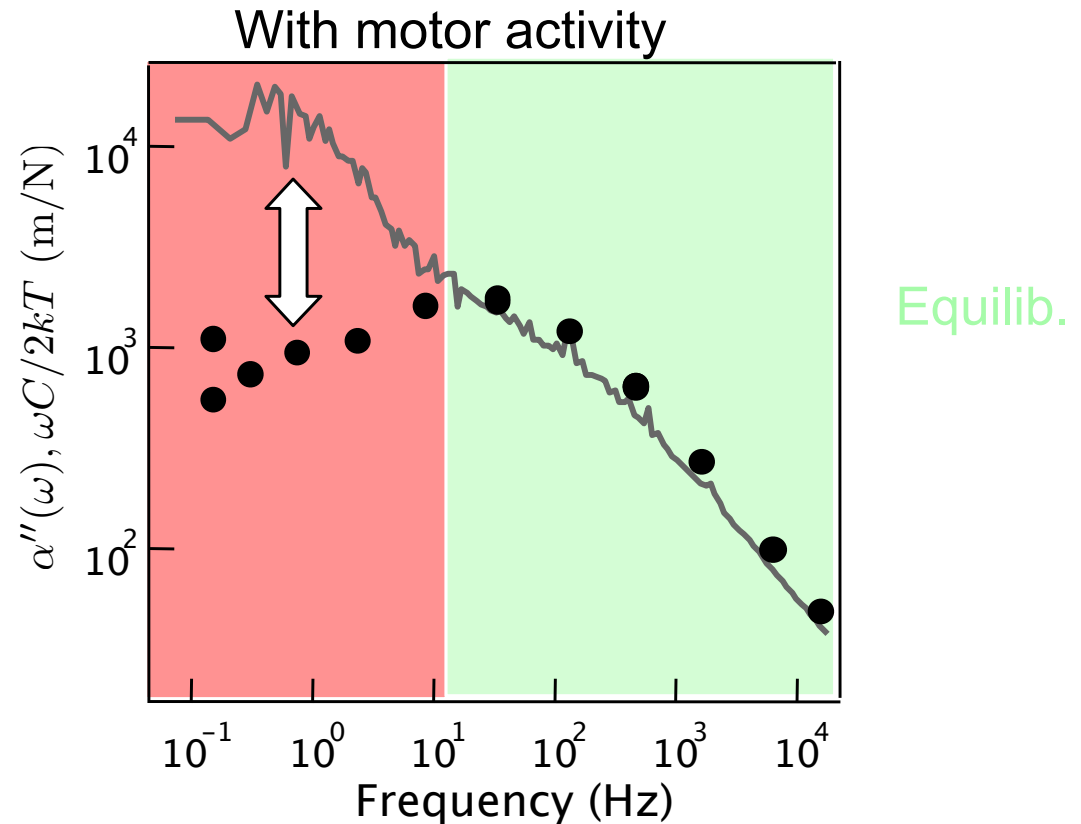
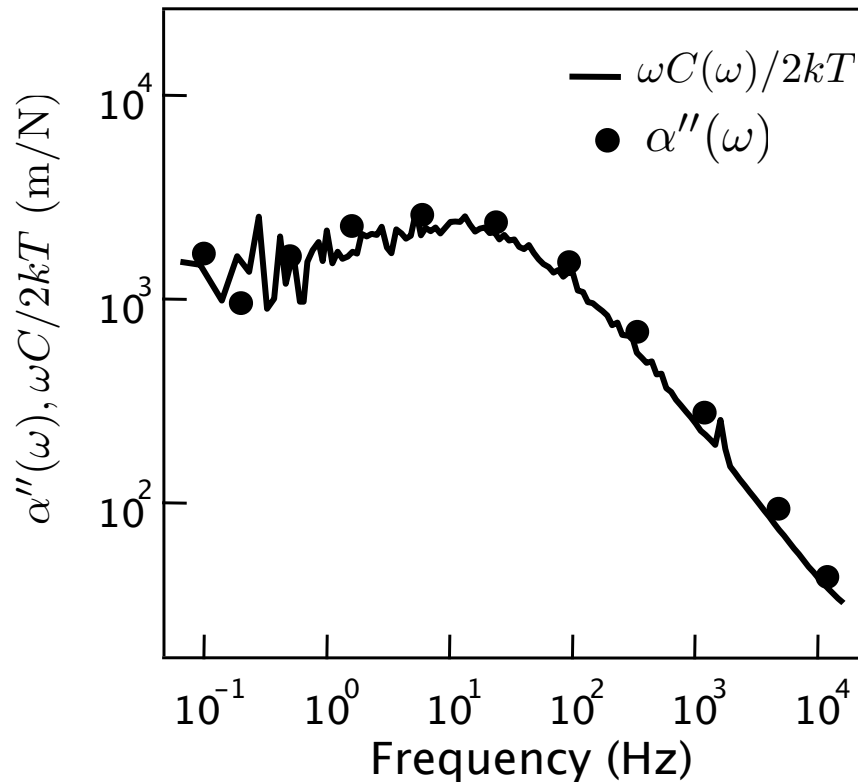
Simultaneous measurement of *passive* fluctuations and **active** response to calib. forces (Mizuno, et al. 2007)

$$C(\omega) = \int \langle x(t)x(0) \rangle e^{i\omega t} dt = \frac{2kT}{\omega} \alpha''(\omega)$$

$$x_\omega = \alpha(\omega) f_\omega$$

A red arrow points from the text 'Simultaneous measurement of passive fluctuations and active response to calib. forces (Mizuno, et al. 2007)' to the $\alpha(\omega)$ term in the equation above.

Effect of molecular motors: *Active Gels*

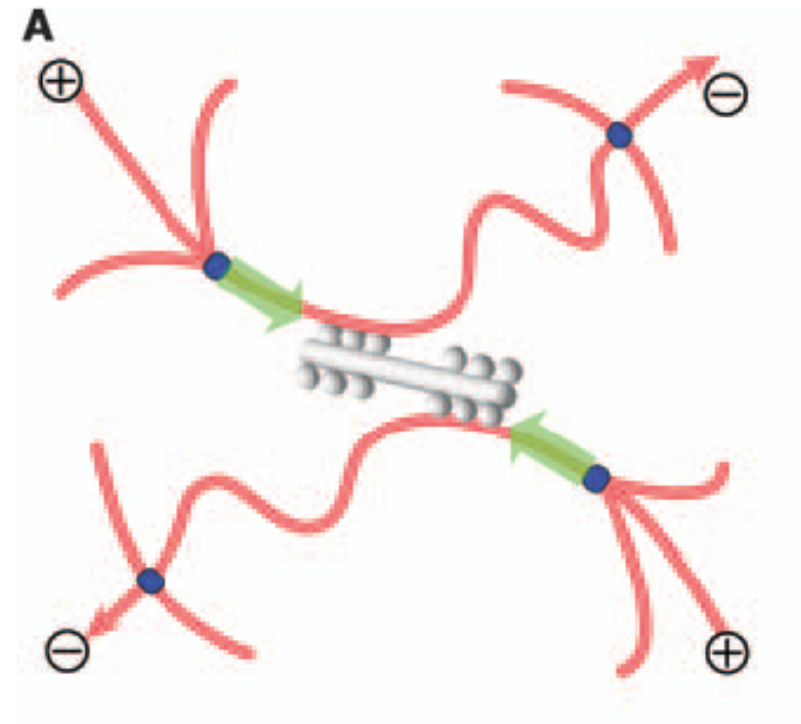
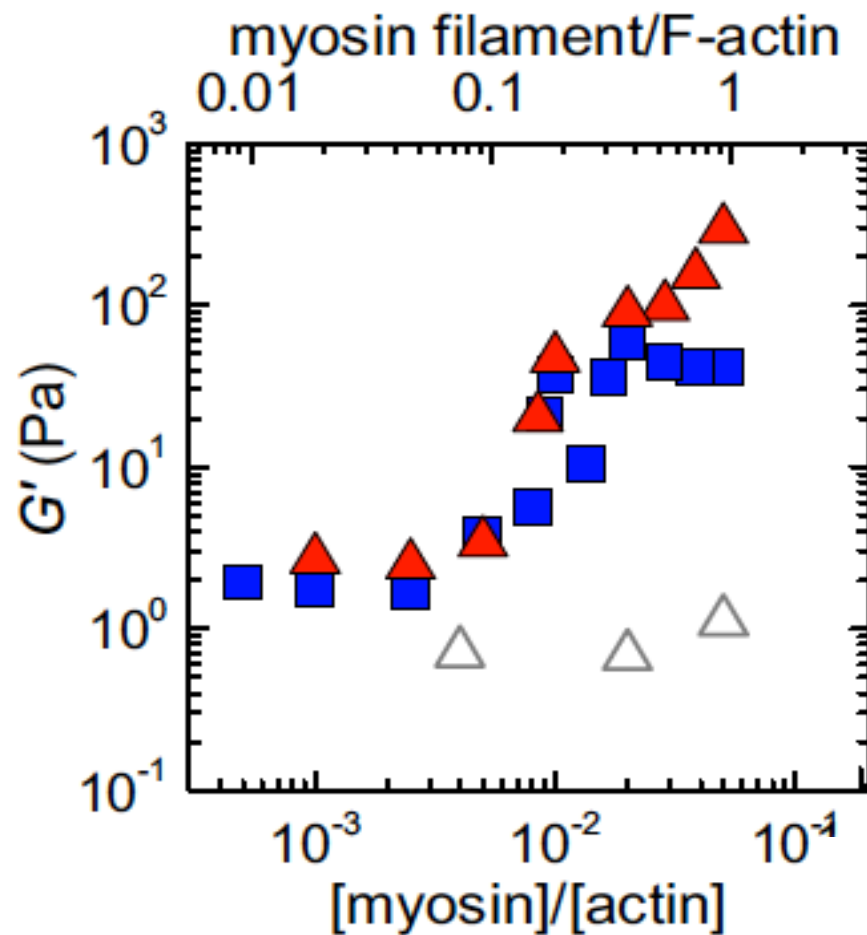


$$C(\omega) = \int \langle x(t)x(0) \rangle e^{i\omega t} dt \neq \frac{2kT}{\omega} \alpha''(\omega)$$

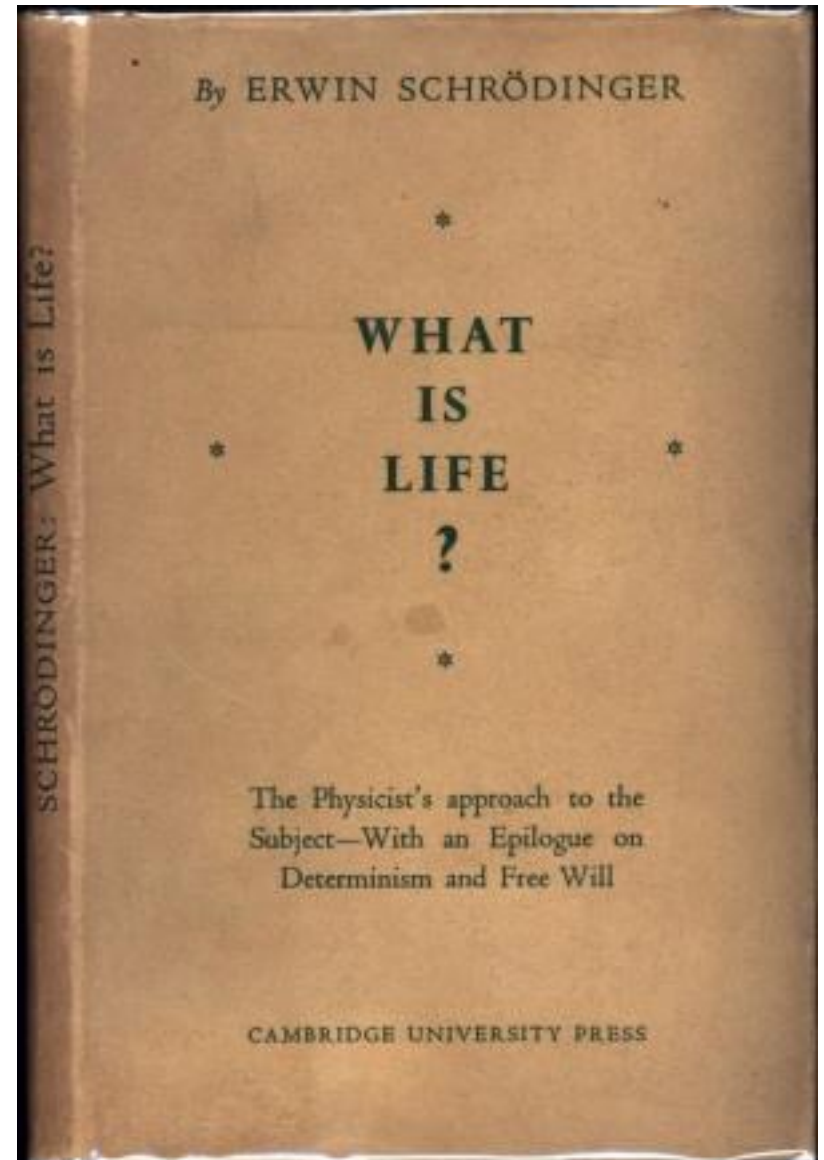
Mizuno, Tardin, Schmidt,
FCM, *Science* 2007.

Contractile forces stiffen network

Koenderink et al.,
PNAS, **106**:15192 (2009).



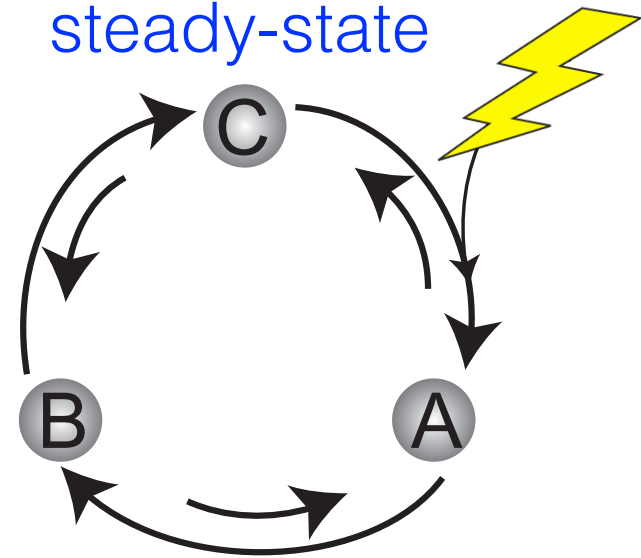
Living systems must evade equilibrium (Schrödinger 1944)



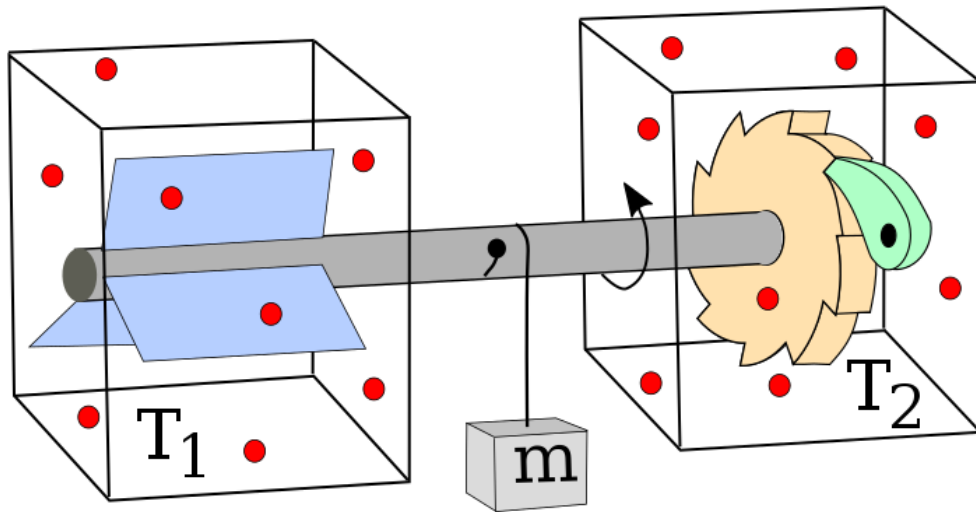
Living systems must evade equilibrium (Schrödinger 1944)



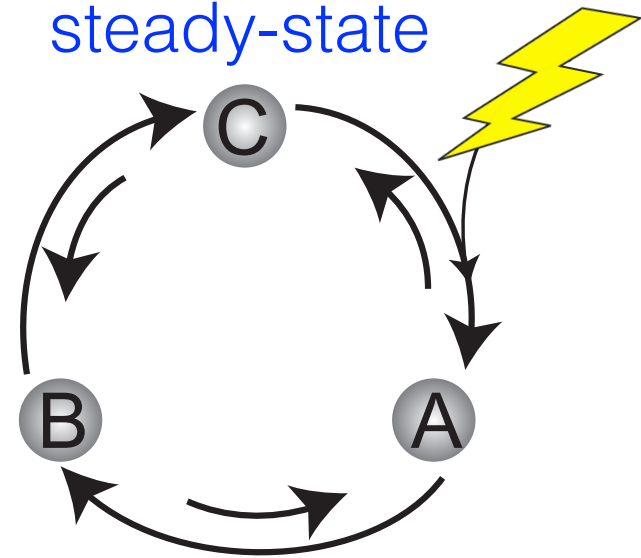
Non-equilibrium
steady-state



Living systems must evade equilibrium (Schrödinger 1944)

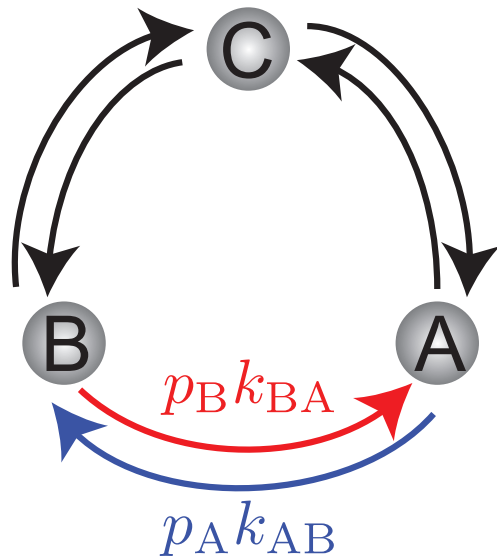


Non-equilibrium
steady-state



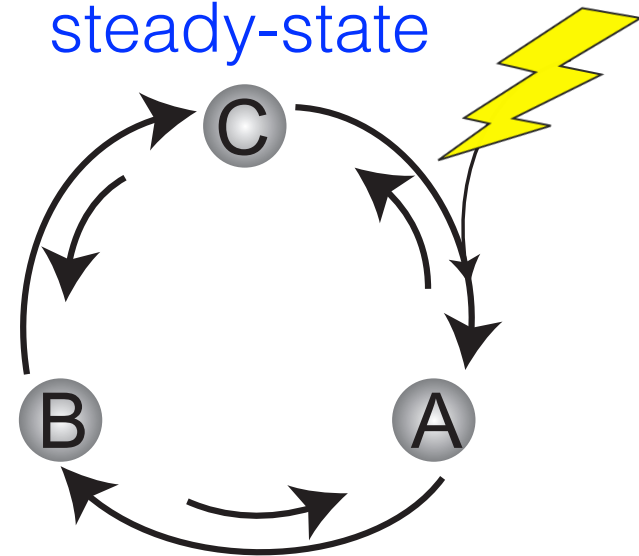
Living systems must evade equilibrium (Schrödinger 1944)

Equilibrium dynamics



Currents J and cycles
forbidden in equilibrium

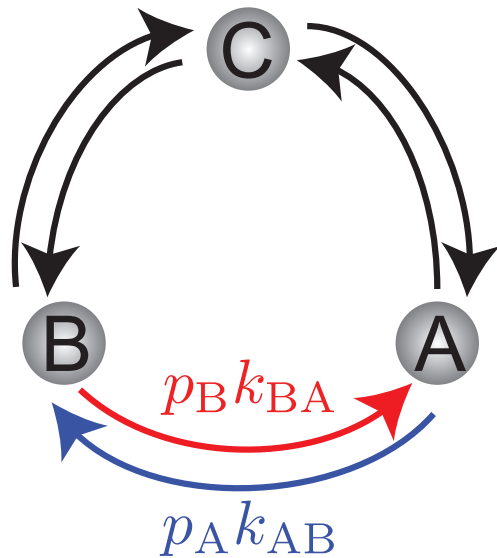
Non-equilibrium
steady-state



$$J_{AB} = p_A k_{AB} - p_B k_{BA}$$

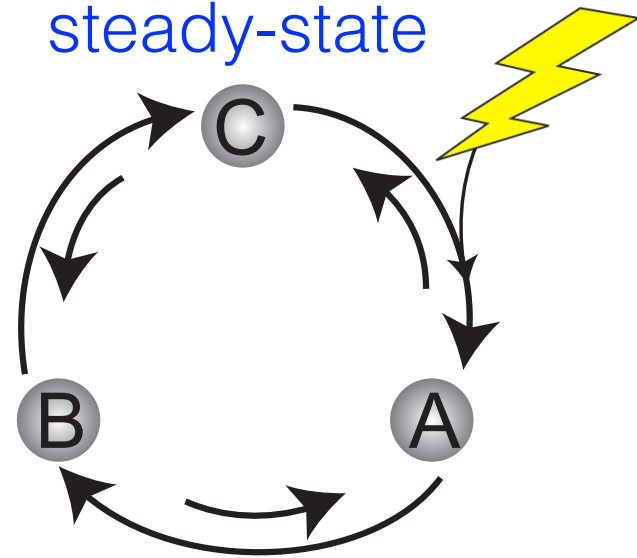
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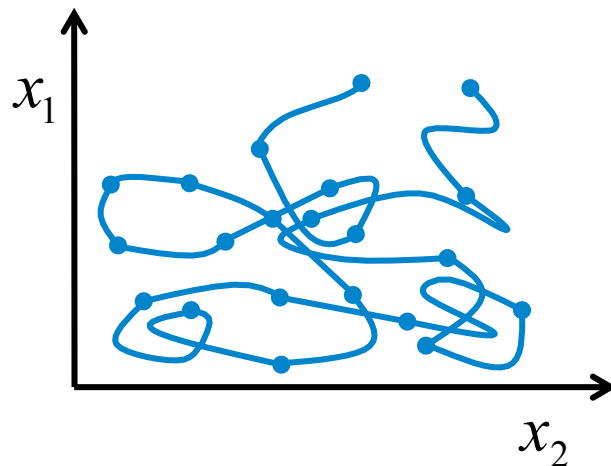
Non-equilibrium
steady-state



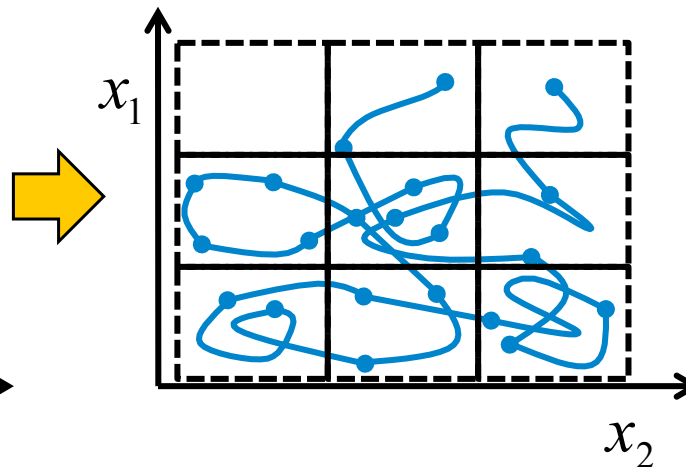
$$\frac{\partial}{\partial t} \rho = -\nabla \cdot J = 0$$

Can we see Broken Detailed Balance at macro-/meso-scale?

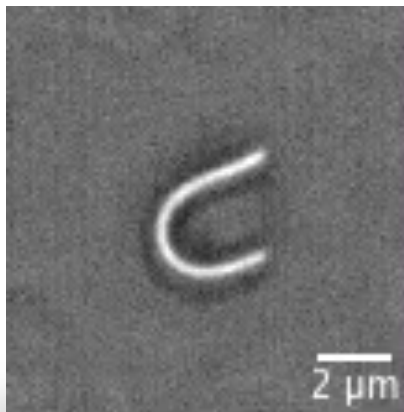
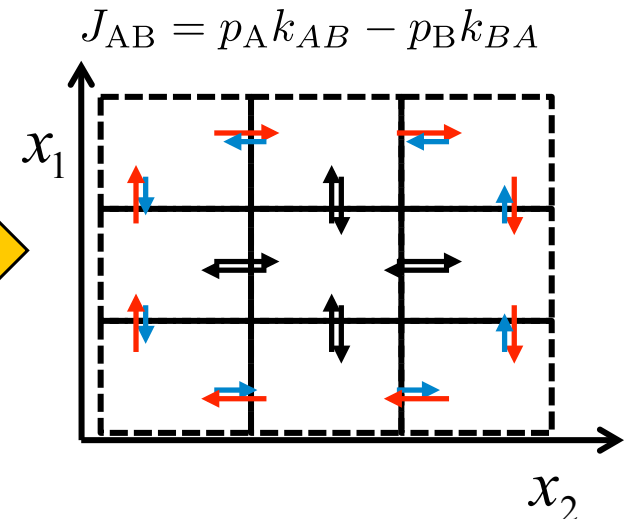
Trajectories in continuous configurational phase space



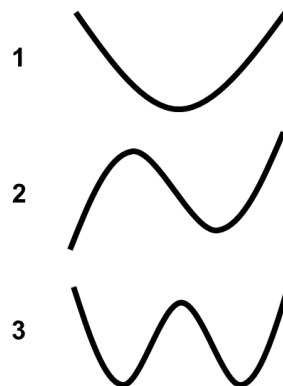
Coarse grain phase space



Probability fluxes



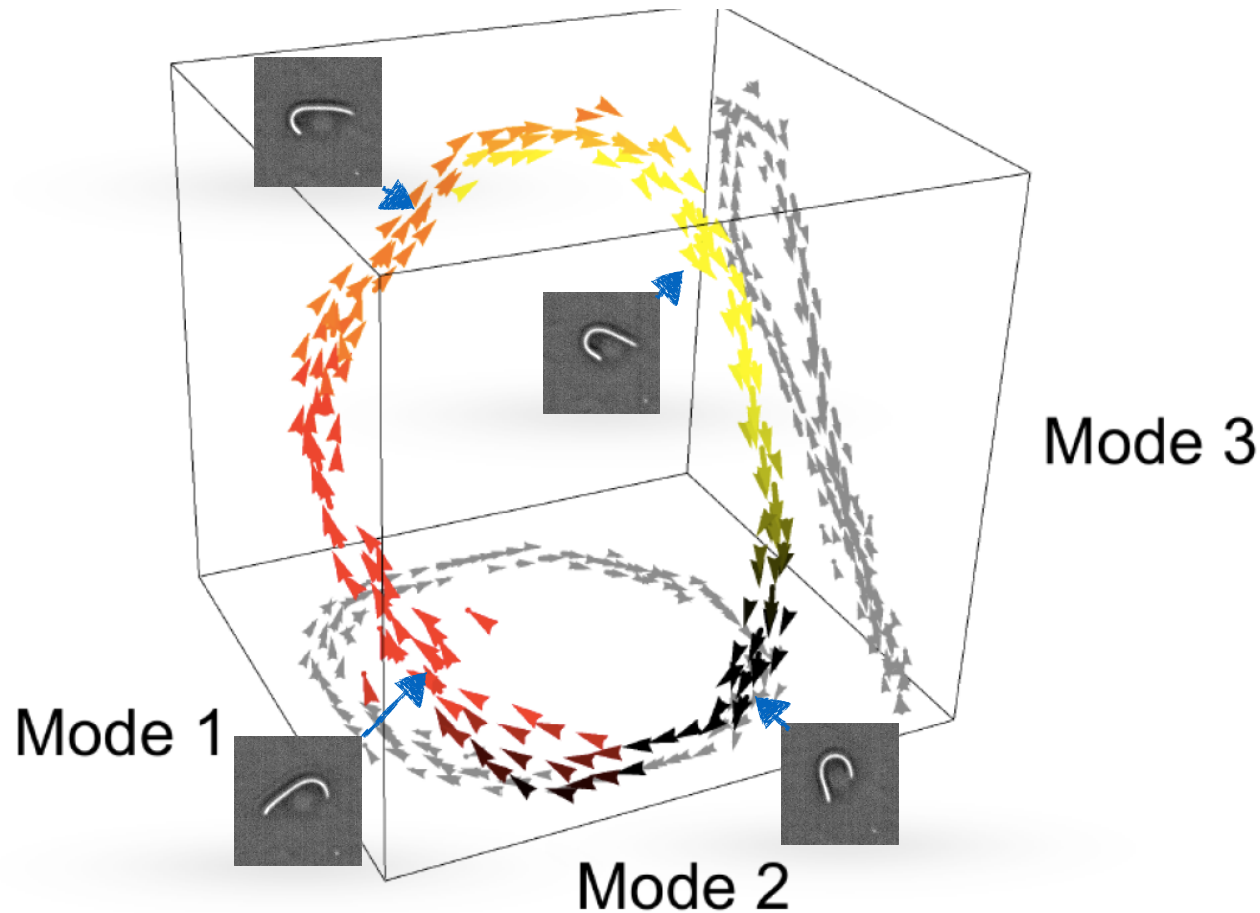
Normal modes



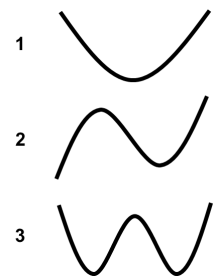
Chlamydomonas flagellum

Battle, Broedersz, Fakhri, Geyer,
Howard, Schmidt, FCM, *Science* 2016.

Projected current/flux in configurational phase space



Normal modes



See also:

Ma, Klindt, Riedel-Kruse, Jülicher, and Friedrich, PRL **113**, 048101 (2014)

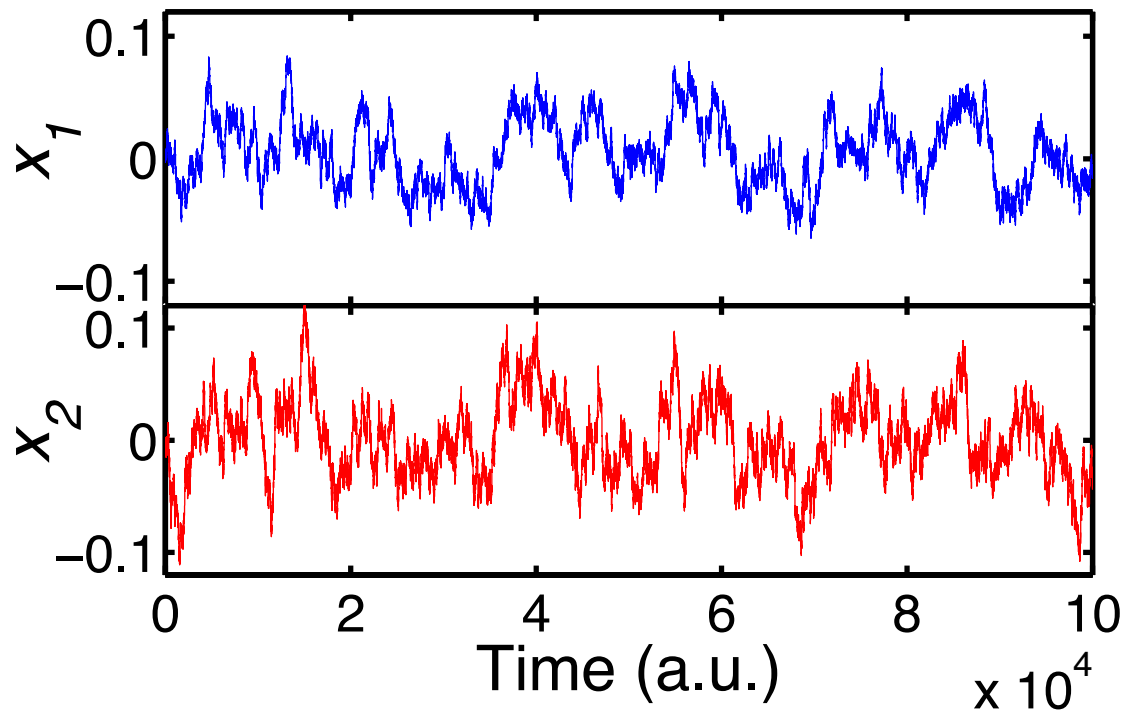
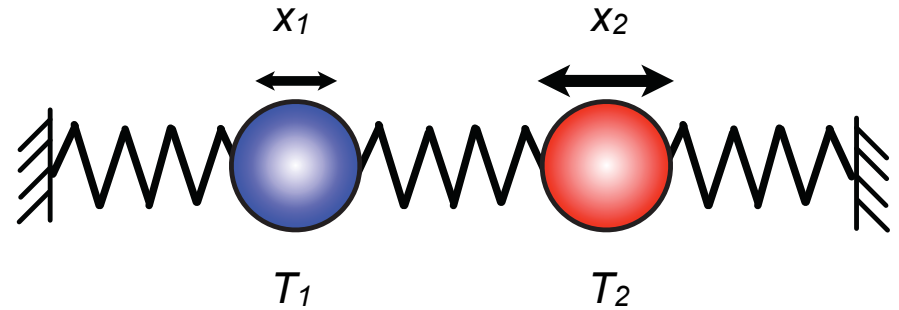
Geyer, Jülicher, Howard, and Friedrich PNAS, 110 (45), 18058(6) (2013)

Coupled beads: A toy model

$$\frac{dx_1}{dt}(t) = k(x_2(t) - 2x_1(t)) + \sqrt{T_1}\xi_1(t)$$

$$\frac{dx_2}{dt}(t) = k(x_1(t) - 2x_2(t)) + \sqrt{T_2}\xi_2(t)$$

$$\langle \xi_i(t) \rangle = 0 \quad \langle \xi_i(t') \xi_j(t) \rangle = 2\delta_{i,j}\delta(t - t')$$

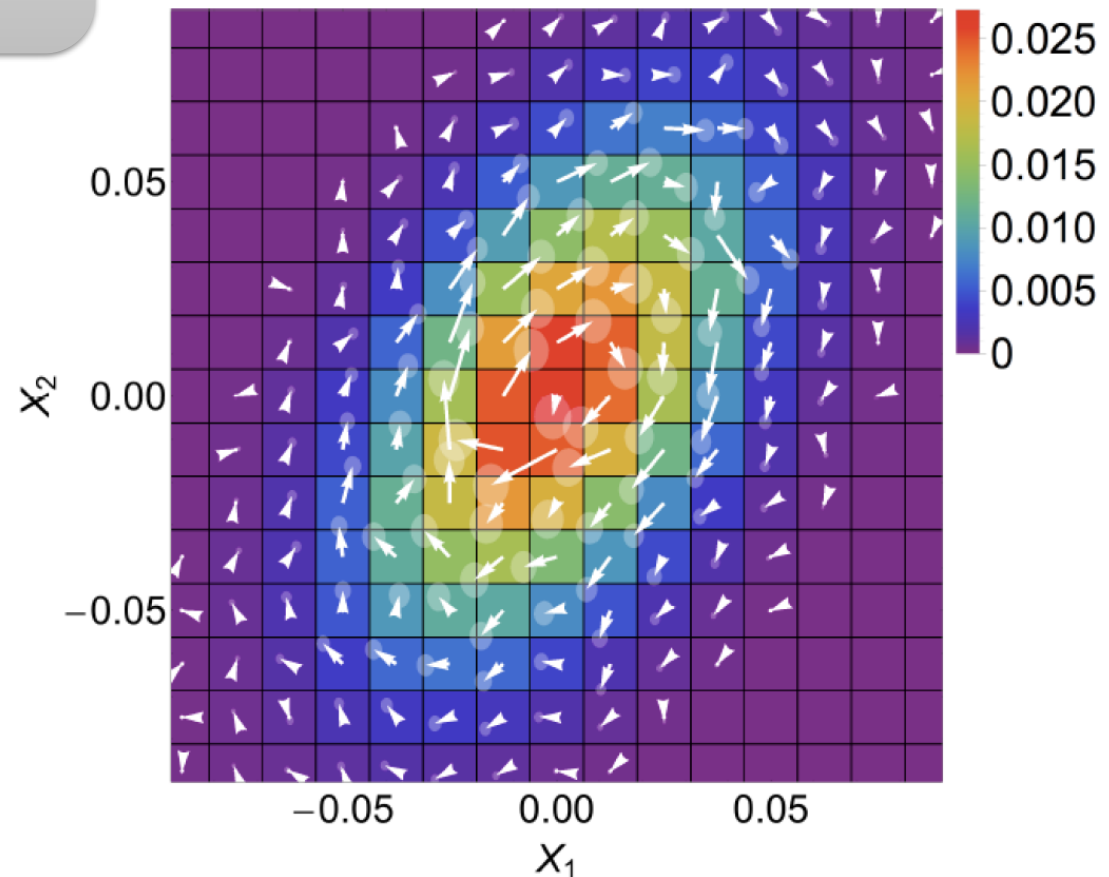
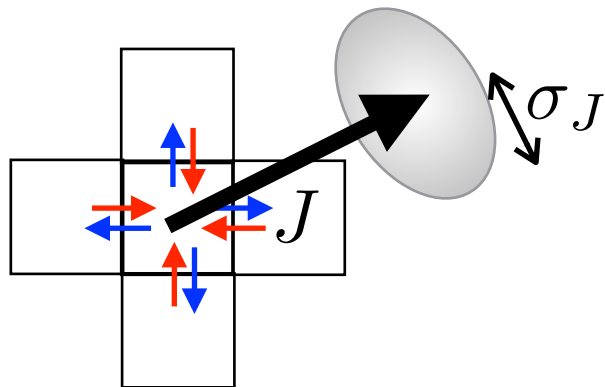
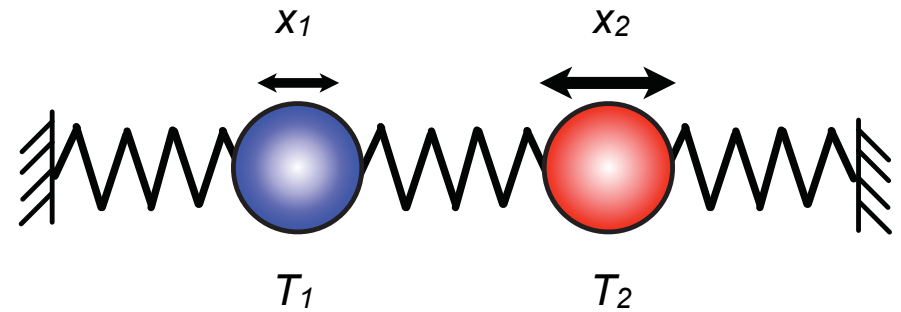


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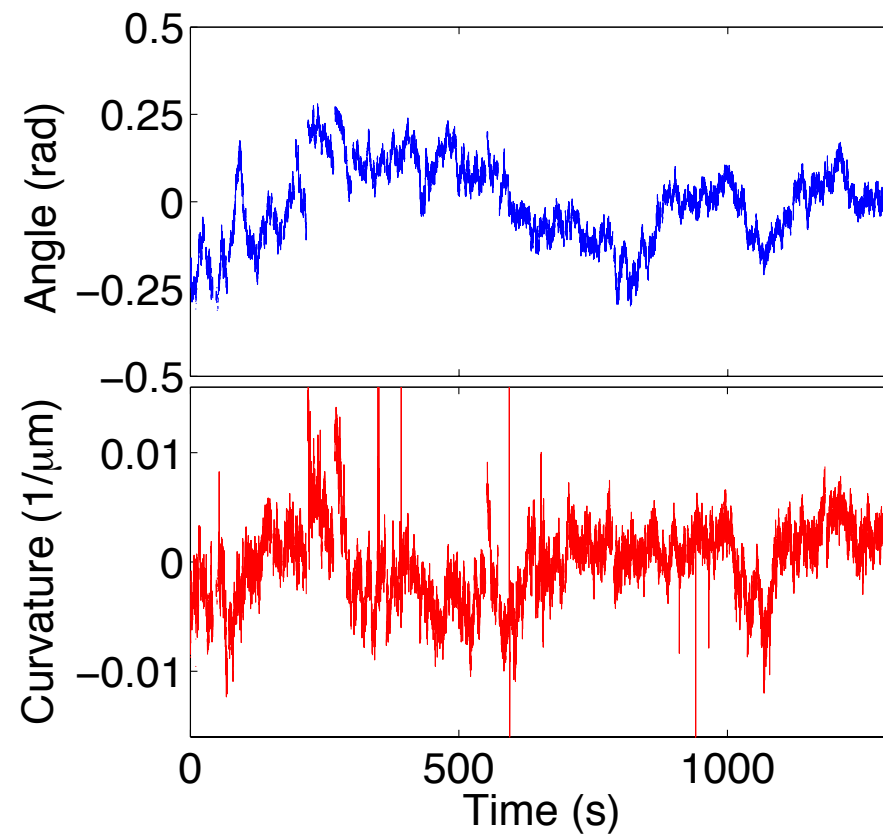
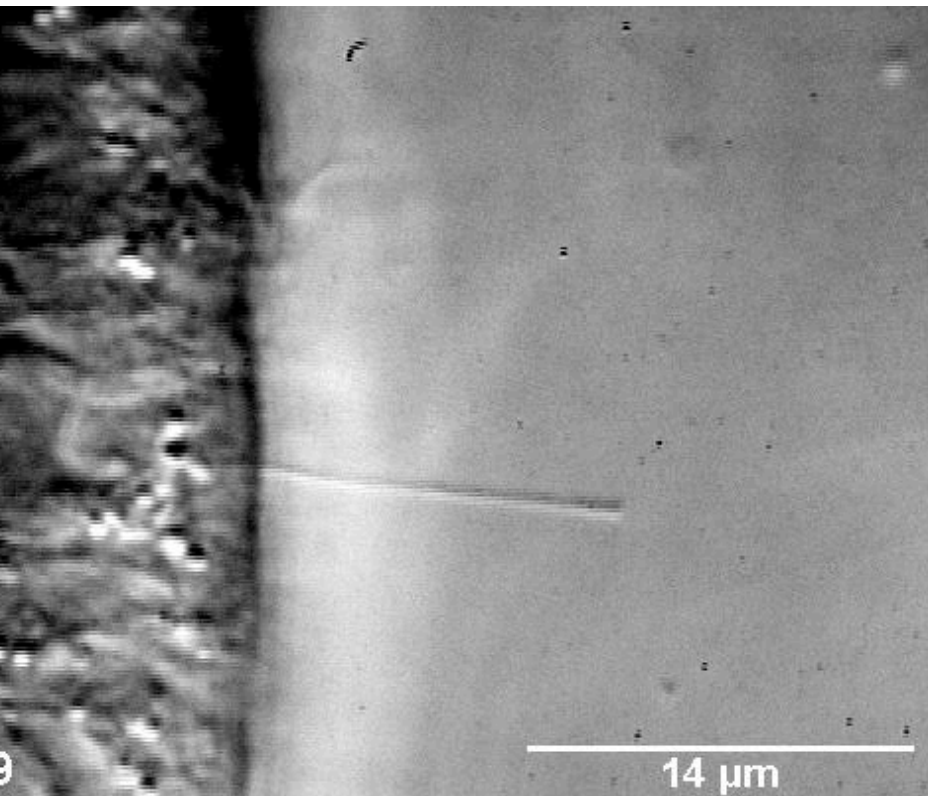
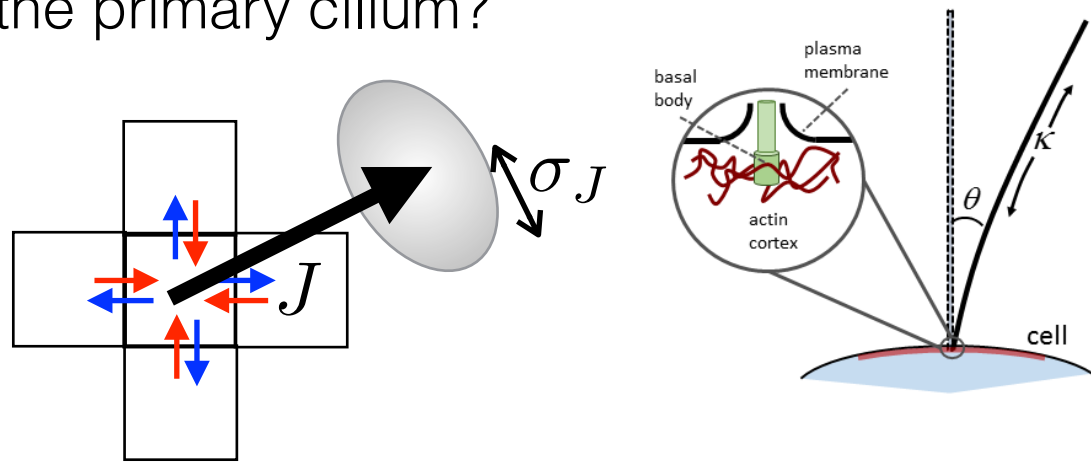
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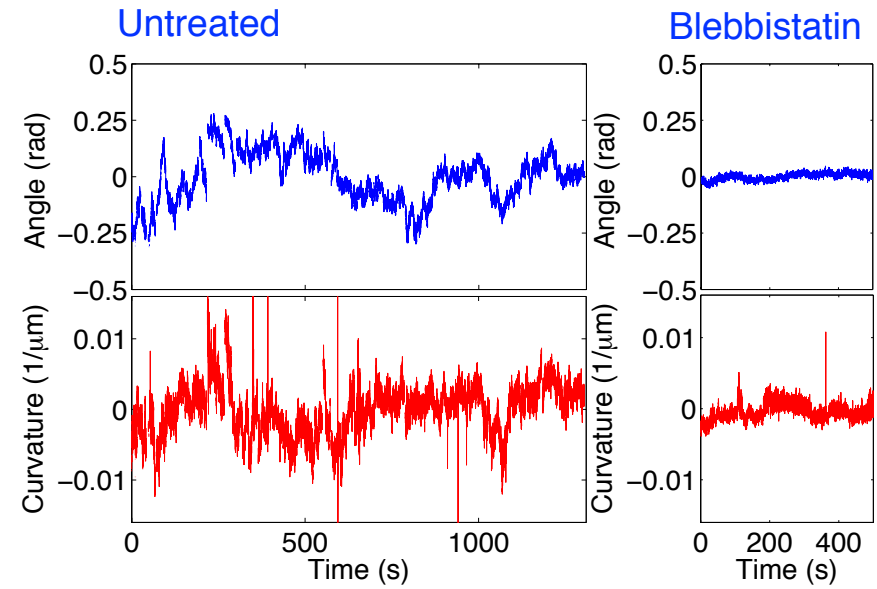
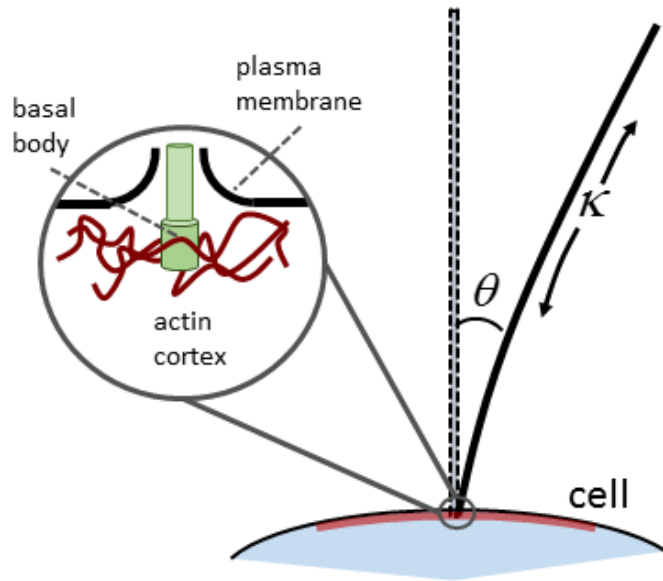
see also: Grosberg & Joanny 2015.

What drives the dynamics of the primary cilium?

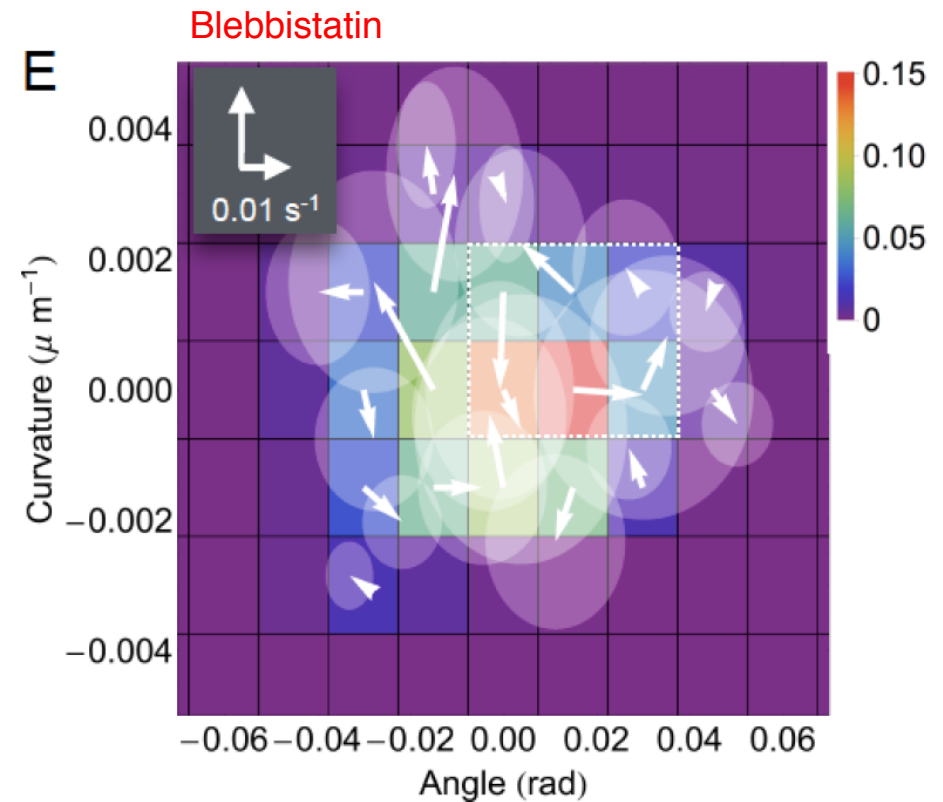
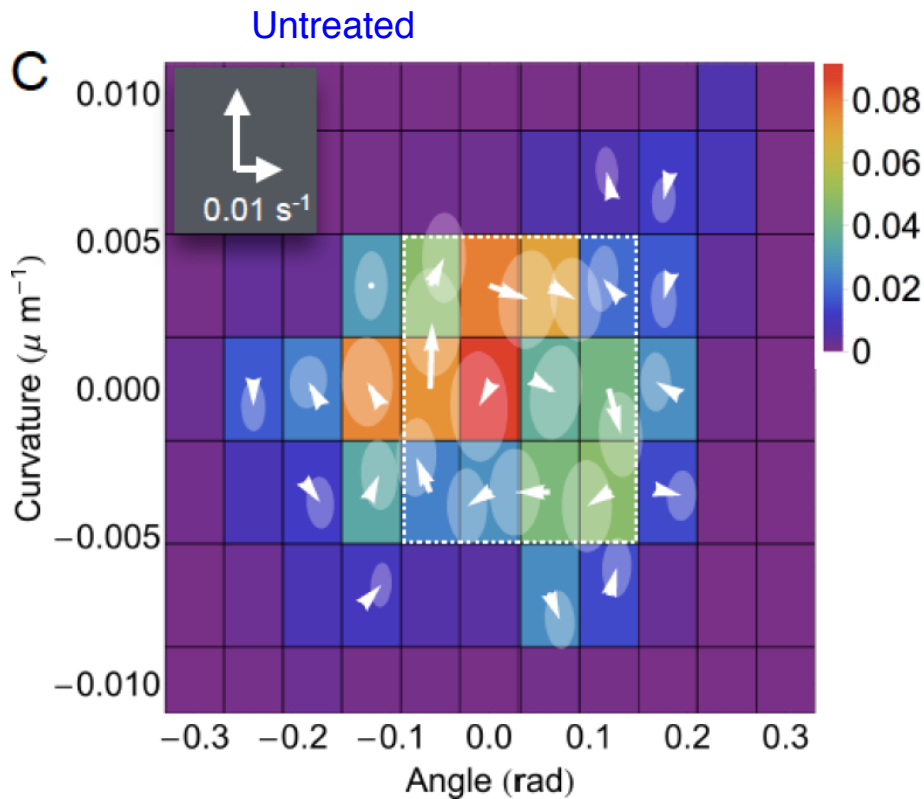
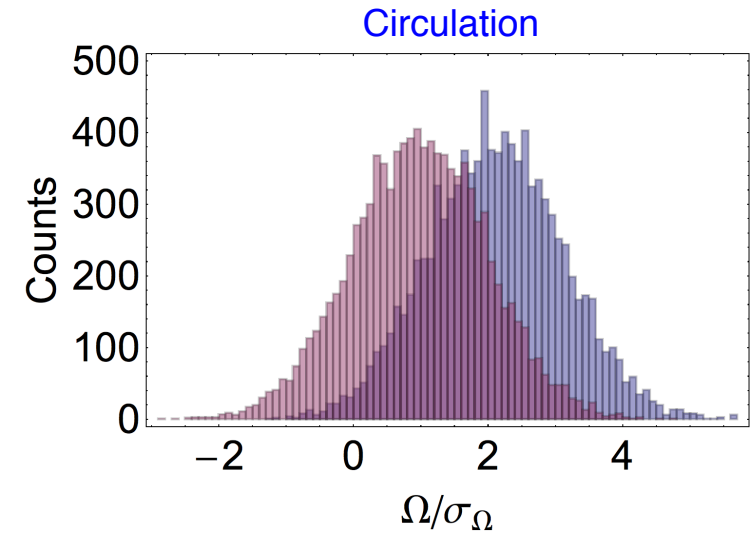
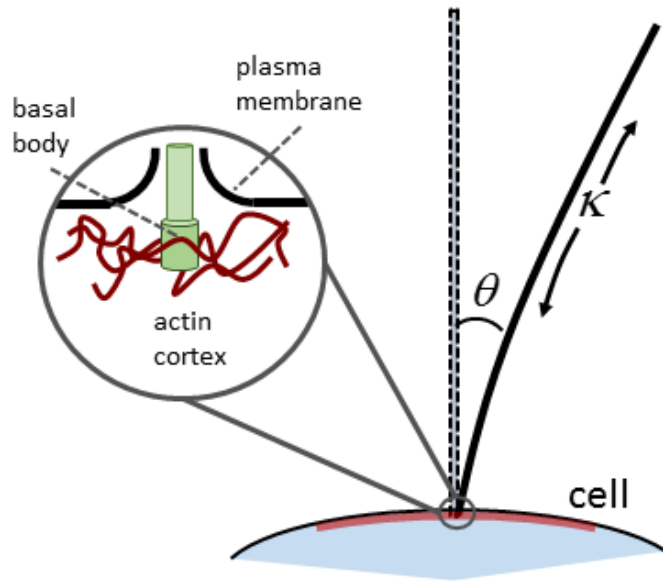


Chris Battle (University of Göttingen)

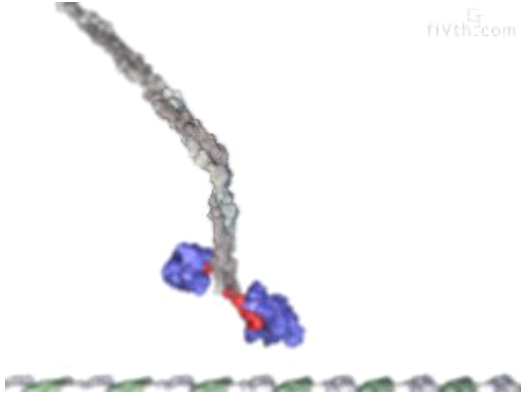
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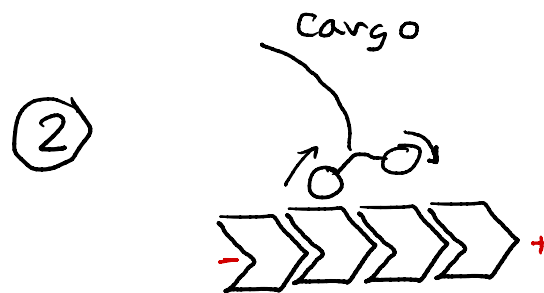
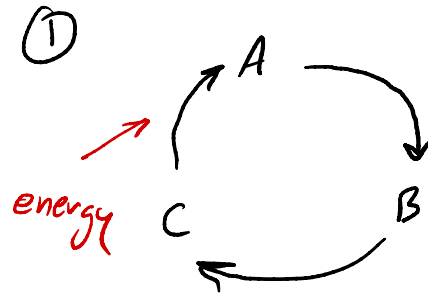


Molecular motors: minimal ingredients



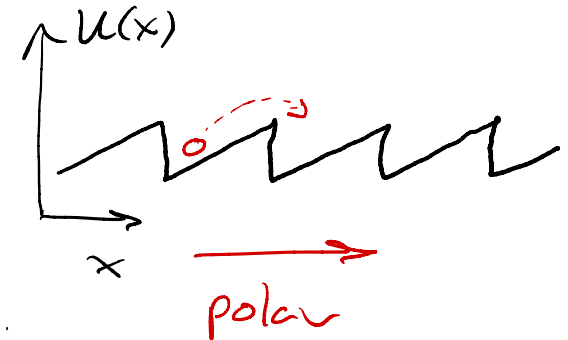
Vale lab, UCSF

Broken Detailed Balance, or
broken t -reversal symm.



broken spatial symm.
a polar filament

Schematically,



see,
Magnasco, PRL 71: 1477 (1993); Prost et al. PRL 72: 2652 (1994).

All known motors operate on polar substrates

Note that actin and microtubules are both polar, which is essential for *myosin* and kinesin/dynein function.

There are no known motors on *intermediate filaments* or *septin filaments*, which are apolar.

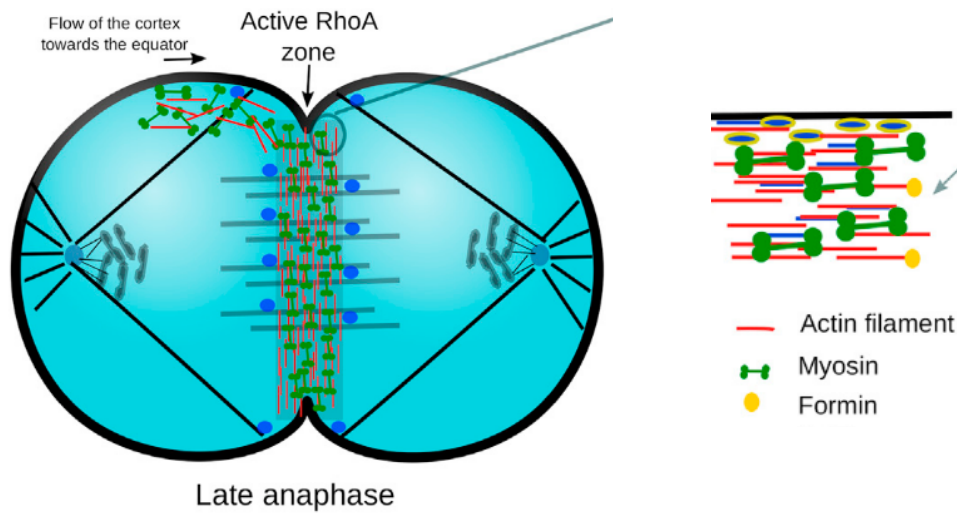
But, there is evidence for contractility without acto-myosin:

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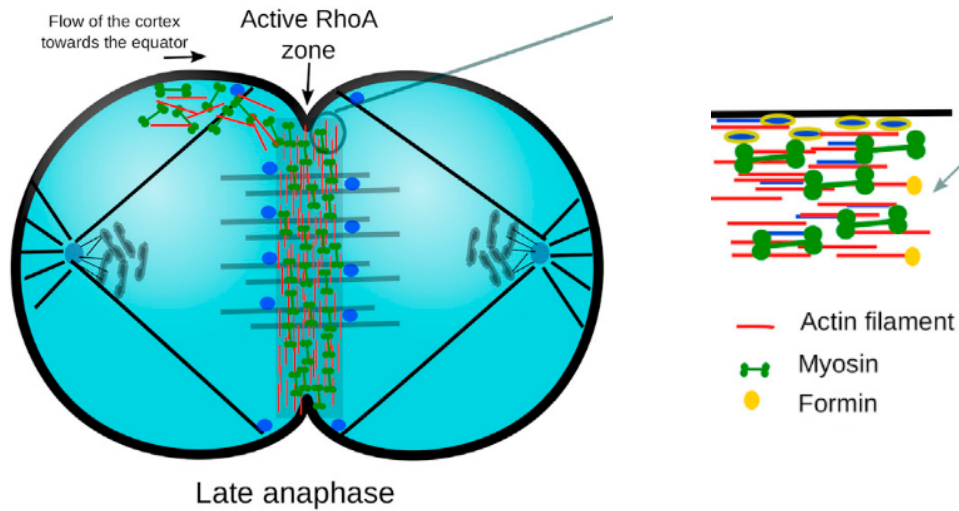


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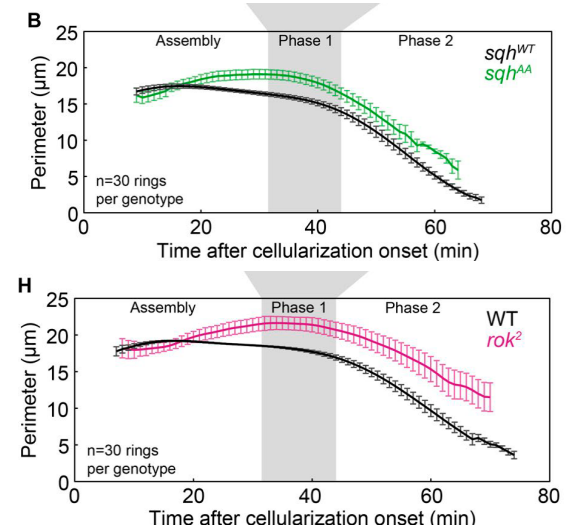
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Schwayer, C. et. al. *Developmental Cell* (2016)

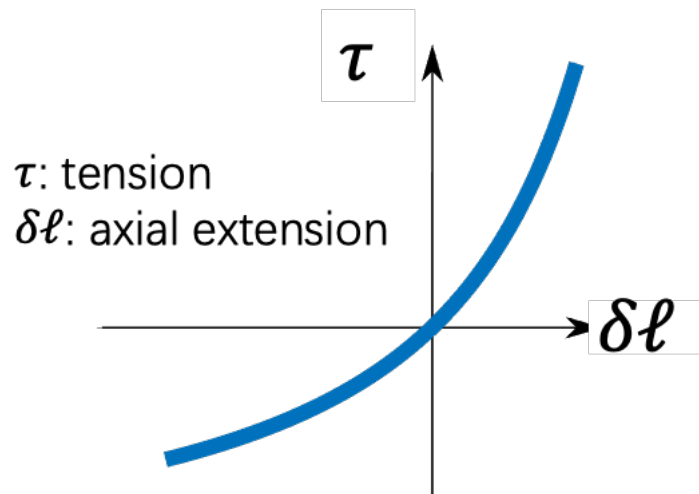


Xue, Z., & Sokac, A. M., *J Cell Biol* (2016)

Black: With Motor Activity
Red/green: Without Motor Activity

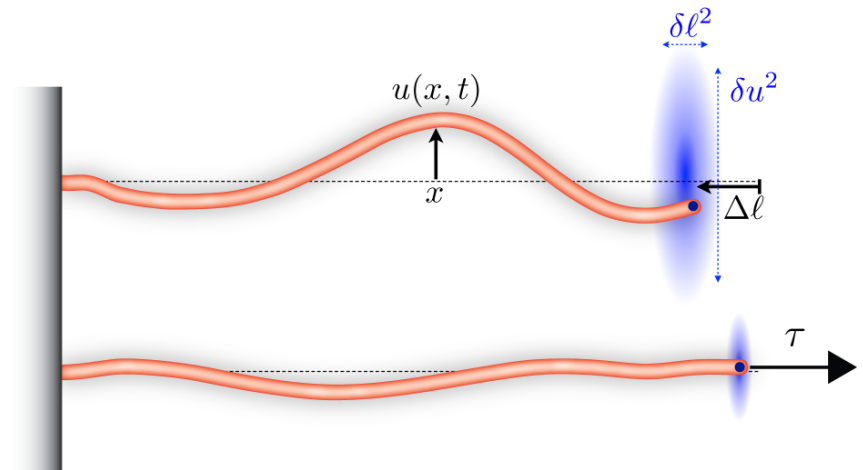
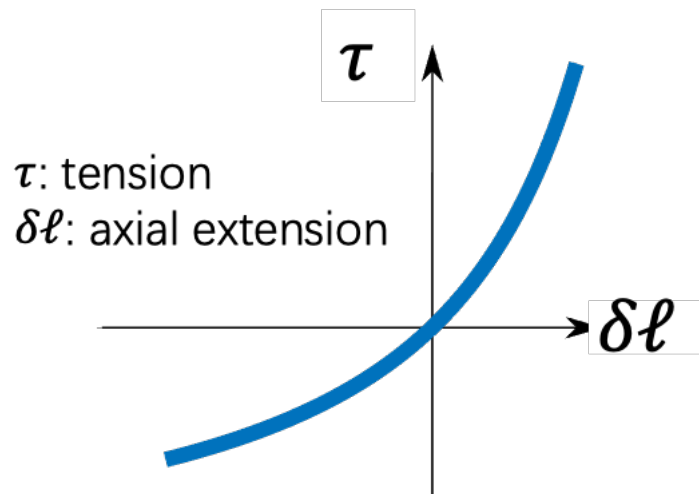
Another broken symmetry: force-extension

The nonlinear force-extension is asymmetric under extension/compression:



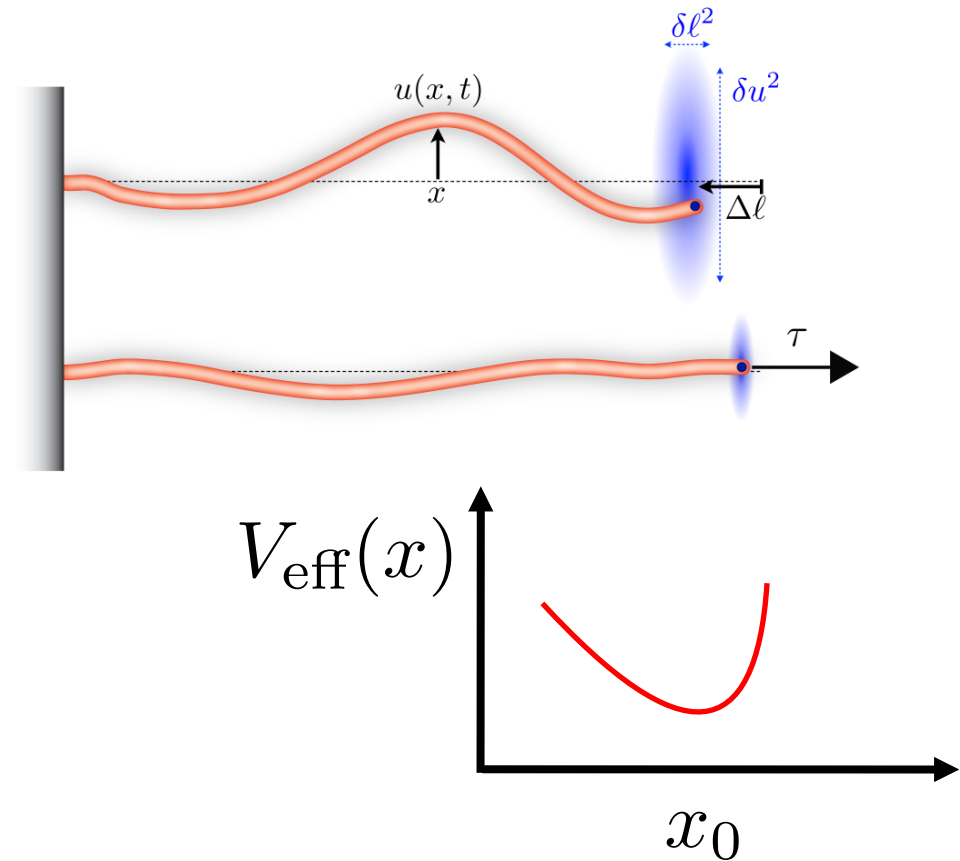
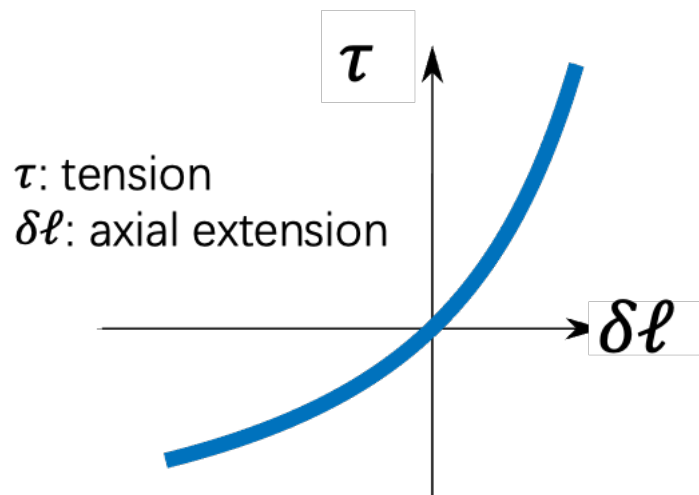
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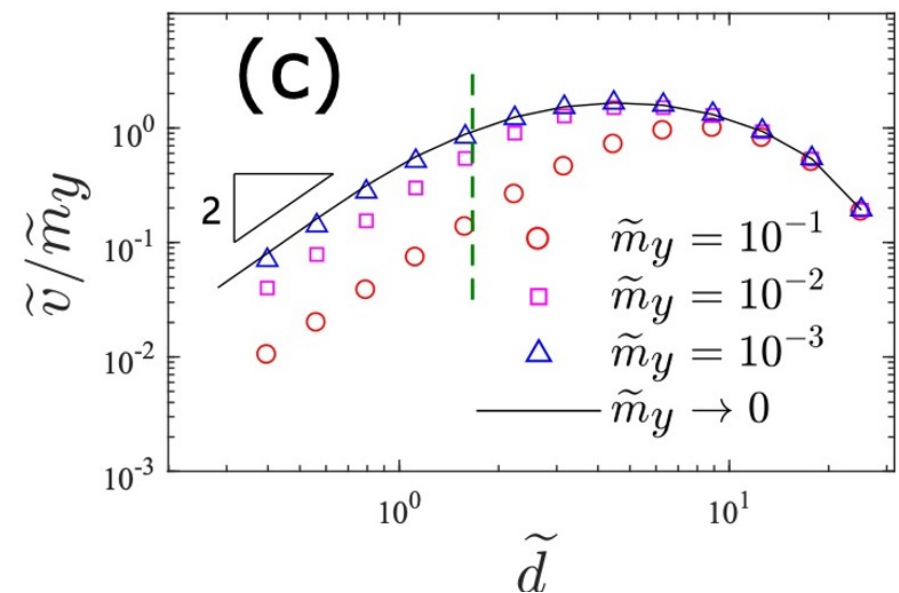
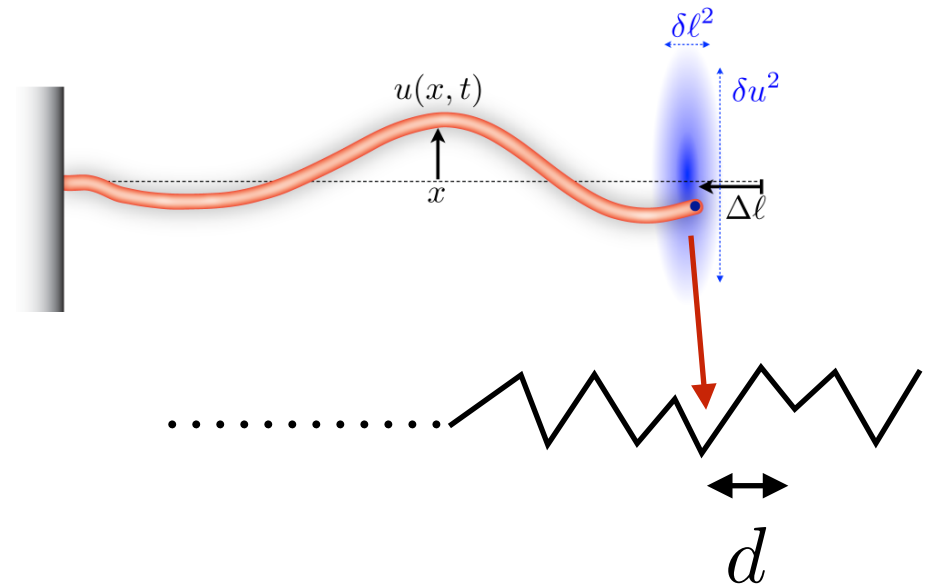
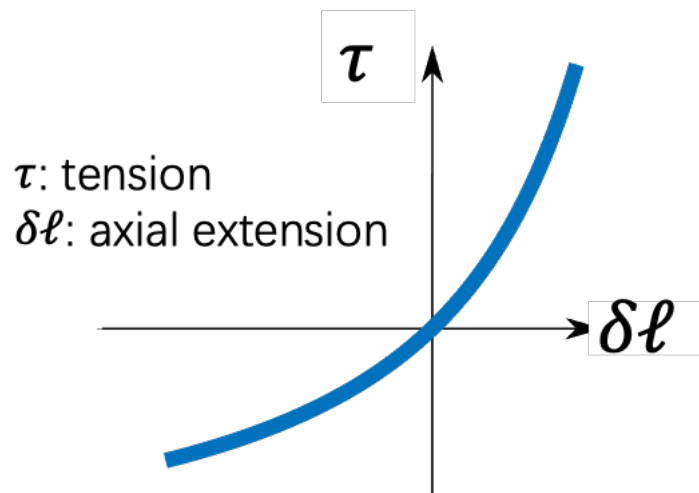
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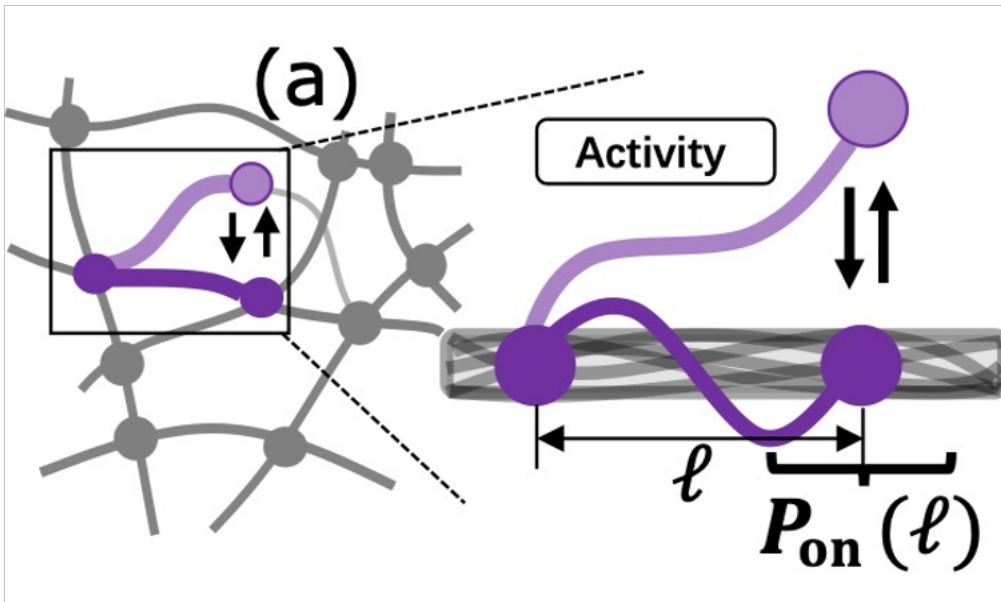


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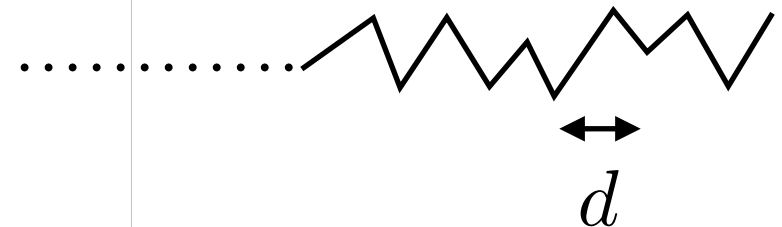
The nonlinear force-extension is asymmetric under extension/compression:



Minimal Model

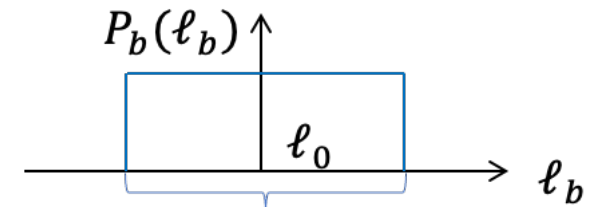


Rugged binding landscape
with typical spacing d



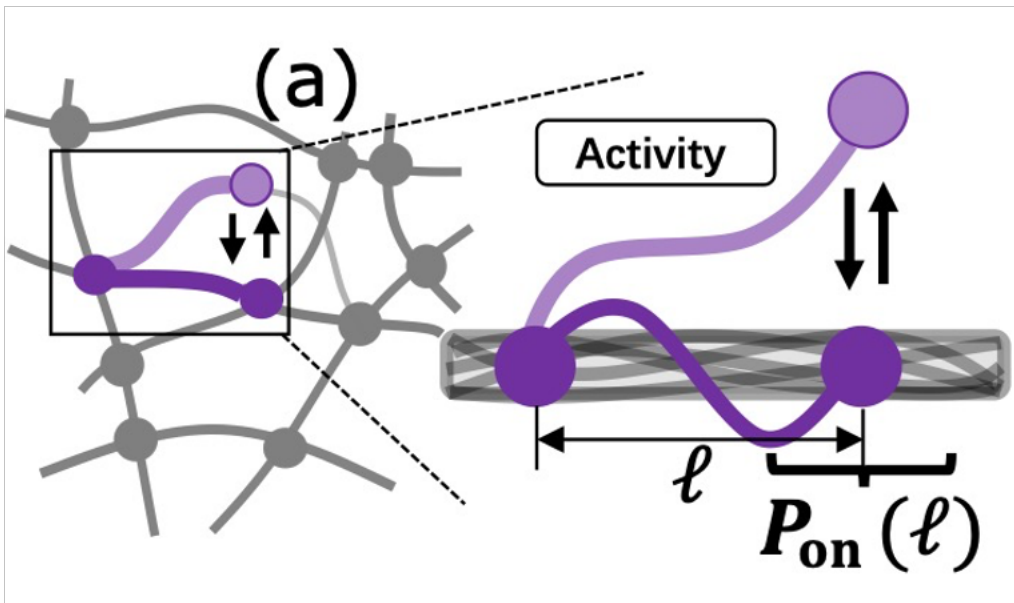
➤ Constant Transition Rates ω_{on} , ω_{off}
(breaking detailed balance)

- Two States: On and Off
- Active Crosslinkers (Not Motors)
- $\tau(\ell)$: Asymmetric force-extension
- $P_{on}(\ell)$: Steady State Distribution in On state



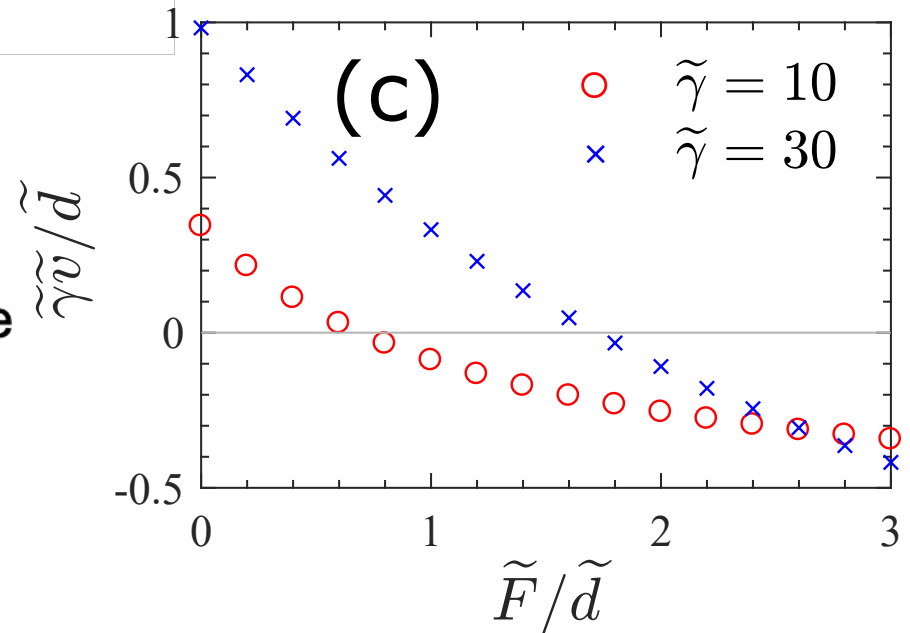
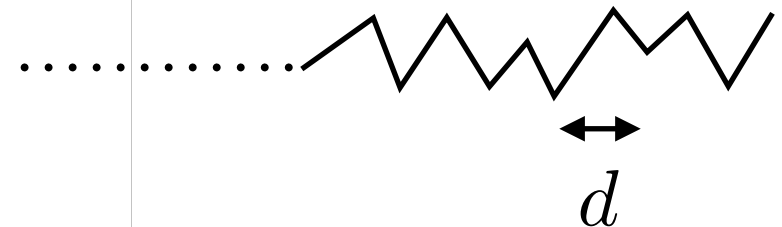
P_b : symmetric or asymmetric
 ℓ_0 : rest length

Minimal Model



- Two States: On and Off
- Active Crosslinkers (Not Motors)
- $\tau(\ell)$: Asymmetric force-extension
- $P_{on}(\ell)$: Steady State Distribution in On state

Rugged binding landscape with typical spacing d



Active fluctuations, Broken Detailed Balance & Contractile Forces

- Interest in synthetic active solids inspired by cytoskeleton
- We propose a non-motor mechanism for contractile force generation
- Also, very interesting critical/phase transition phenomena governed by contractility
- **Collaborators:** Schmidt, Koenderink, Weitz, Fakhri, Guo, Mizuno, Alvarado, Chen, Markovich, Broedersz, Sharma, Sheinman, ...

Reviews:

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